

THE FRITSMAS FACTOR

Your
Interactive
Hemostasis
Resource

Managing Hemophilia Therapy Rasputin to Recombinants

George A. Fritsma, MS MLS

The Fritsma Factor; Your Interactive Hemostasis Resource

www.fritsmafactor.com

Hemophilia case study

Hemophilia in history

Alexis and Rasputin

AIDs and recombinants

Extended half-life recombinants

Gene transfer



The Fritsma Factor



40 YO Af-Am Hemophilic Male

- Bleeding into ankle at midnight
 - Anatomic soft-tissue and joint bleeds
 - Tried RICE, DDAVP (Stimate®) inhaler
- Ran out of factor VIII (FVIII) concentrate
 - Subsidized by Medicare and Medicaid through ~100 national hemophilia centers
 - National Hemophilia Foundation: www.hemophilia.org
- Night tech and on call clin path resident
 - Determine residual patient factor VIII activity
 - Compute FVIII concentrate dosage
 - Order FVIII concentrate from transfusion service
 - Reconstitute with sterile water, administer as IV push
 - Subsequently determine therapeutic factor VIII activity



Calculating FVIII Concentrate Dosage

- One unit = activity in 1 mL normal plasma or 100%
- Determine plasma volume based on patient's weight
 - Blood volume (BV, mL) = weight (kg) x 70 mL/kg
 - Use 60 mL/kg for obese, BMI 25–30
 - Use 50 mL/kg for morbidly obese, BMI >30
 - Plasma volume (PV, mL) = BV x (100%–HCT%)
- Decide on units (%) of FVIII concentrate desired

$$\text{Units of FVIII required} = (\text{desired FVIII in units/mL} - \text{initial units/mL}) \times \text{PV (mL)}$$

If Factor Assay not Available

- When timing is critical, assume <1% activity or...
- Estimate residual FVIII from PTT

Factor VIII	PTT
40%	35 s
30%	50 s
20%	65 s
10%	90 s
1%	120 s
<i>Example only, do not use</i>	



- Collect baseline plasma
 - Assay or freeze and confirm with a real assay during next day shift
- Maintain patient database

40 YO Af-Am Hemophilic Compute FVIII Dosage

- Weight is 80 kg, HCT 40%, factor level <1%
- Determine plasma volume
 - Blood volume (mL) = 80 kg x 70 mL/kg = 5600 mL
 - Plasma volume (mL) = blood volume (5600 mL) × (100%–40%) = 60% × 5600 = 3360 mL
- Determine units of FVIII required:
 - Choose to reach 80% FVIII (0.8 units/mL), therefore...
 - Units of FVIII required = (0.8 units/mL–0) x 3360 mL = 2688 (2700) units
 - Typical concentrate vial provides ~1000 units, use 3
- Avoid overdose: potentially thrombotic and wasteful

Plasma-derived FVIII Concentrates

- Theoretical risk of HBV, HCV, HIV transmission
 - Hemofil-M[®], Monarc-M[®](Baxter), Monoclate-P[®]
 - Human and animal plasma matrix
 - Purification: immunoaffinity column, solvent-detergent, Pasteurization, viral filtration, combinations
- Seroconversions per CDC surveillance: 0
 - Data from 2003; predicted risk, 1:60,000
- <25% of FVIII concentrates used in industrialized countries



The Fritsma Factor

NDC 0053-7656-01
One vial with diluent

LOW

Monoclate-P[®] Antihemophilic Factor (Human)

Factor VIII:C Pasteurized
Monoclonal Antibody Purified

For Intravenous Administration Only. Rx only

Storage: Monoclate-P[®] stored in a refrigerator at 2-8°C (36-46°F) is stable for the period indicated by the expiration date on the label. Within this period Monoclate-P[®] may be stored at room temperature not to exceed 25°C (77°F), for up to 6 months. Avoid freezing.

Manufactured by:
CSL Behring LLC
Kankakee, IL 60901 USA
US License No. 1767

CSL Behring

Recombinant FVIII Concentrates

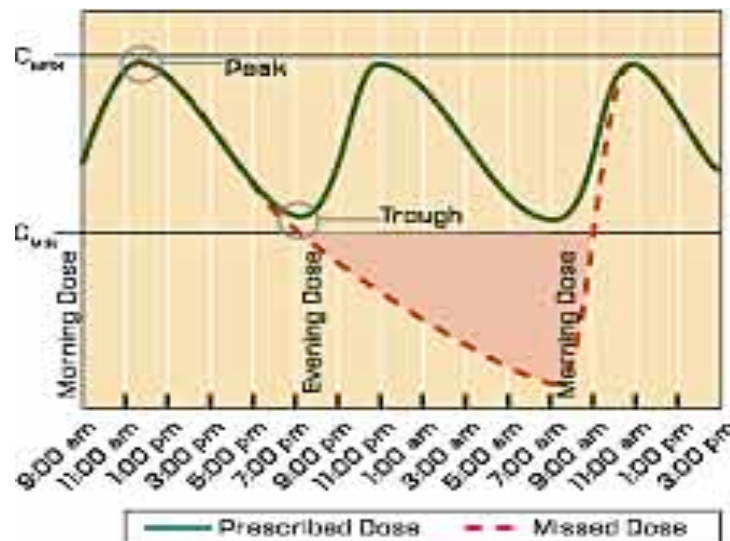
- Serum or albumin in culture medium
 - Helixate, Kogenate, Recombinate
 - Abundant, used for prophylaxis
 - No HBV, HCV, HIV seroconversions
- No protein in culture or preparation
 - No viral risk: Advate
- B-domain-deleted FVIII concentrate
 - Human albumin in culture: ReFacto
 - No protein: Xyntha
 - Extended half-life: Eloctate
 - Caution when using clot-based FVIII assay, use chromogenic FVIII



40 YO Af-Am

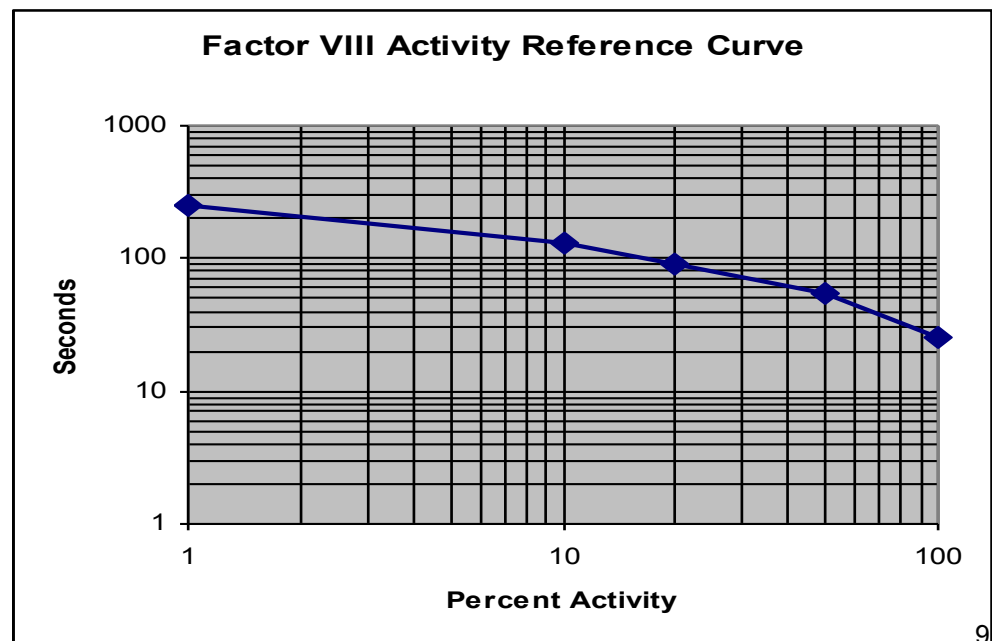
Advate FVIII Concentrate Therapy

- Peak: collected 15 m after administration: 0.3 units/mL
 - Should have been 0.8 units/mL, what happened?
 - Suspect anti-FVIII inhibitor
 - If peak reaches expected value, go to next administration
- Nadir (trough): 12 h after administration
 - Reflects half-life, should reach 50% of desired activity
 - Administer new AHF, use half the dosage second time



One-stage Factor VIII Assay

- Dilute pt plasma and control 1:10 in imidazole buffer
- Add FVIII-depleted reagent plasma 1:1 (FVIII DP)
 - Typical: 100 uL pt plasma dilution/100 uL FVIII-depleted plasma
 - Provides all factors except FVIII
- Add PTT reagent, incubate 3 minutes
- Add CaCl_2 , record interval to clot formation
- Confirm control
- Compare result in seconds to calibration curve to derive plasma activity:



Factor VIII Assay Plasma Dilutions

Parallelism Indicates No Inhibitor

Automated Plasma Dilution	Seconds	Raw Factor VIII Activity	Computed Factor VIII Activity (× dilution)
1:10 “undiluted”	90 s	20%	20%
1:20	105 s	10%	20% (parallel)*
1:40	107 s	5%	20% (parallel)
1:80	110 s	2.5%	20% (parallel)

* <10% difference from undiluted indicates parallelism, no inhibitor

FVIII Assay Plasma Dilutions non-Parallelism Indicates Inhibitor

Plasma Dilution	Seconds	Raw Factor VIII Activity	Computed Factor VIII Activity (× dilution)*
1:10 (undiluted)	80 s	10%	10%
1:20	93 s	8%	16%
1:40	107 s	5%	20%
1:80	108 s	4%	32%

* >10% difference from undiluted = non-parallel & rising, implies inhibitor

- Inhibitor: IgG alloantibody to FVIII concentrate
 - 30% incidence, almost all in severe hemophilia

Kasper CK. Laboratory diagnosis of factor VIII inhibitors. In Kessler C, Garvey MB, Green D, Kasper C, Lusher J. Acquired Hemophilia 2nd Edition. Excerpta Medica 1995

CDC Modified Nijmegen Bethesda Assay

Heat patient and control plasmas 56C/30m & cfg to remove residual FVIII

Make serial dilutions of patient plasma in FVIIIDP

Patient mix

Mix 1 part patient dilution
with 1 part NPP

Control mix

Mix 1 part FVIIIDP with 1
part NPP

Incubate 37C/2h, measure FVIII activity

Patient mix FVIII/control mix FVIII X 100 = % residual activity (RA)

Nijmegen-Bethesda Units (NBU) = $(2 - \log \%RA) / 0.301$

Miller CJ, Platt SJ, Rice AS, et al. Validation of Nijmegen-Bethesda assay modifications to allow inhibitor measurement during replacement therapy and facilitate inhibitor surveillance. J Thrombos

FVIII Inhibitor Therapy: Factor IX Complex

- If ≤ 5 NBU, give high-dose FVIII concentrate
- Prothrombin complex concentrate (PCC, 1980)
 - >5 NBU
 - BaSO_4 extracted human plasma; II, VII, IX, X: Proplex
 - *Activated* PCC: FEIBA, Autoplex
- FEIBA or Autoplex dosage
 - 50 units/kg/12 h standard
 - 70 units/kg/8 h in hemorrhage
 - Limit 200 units/kg/24 h to avoid DIC
 - Cannot monitor: only generalized coag evaluation with PTT



Ludlam DA, Morrison AE, Kessler C.
Treatment of acquired hemophilia. *Semin
Hematol* 1994;31 (Suppl 4) 16-19

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Recombinant FVIIa Concentrate; NovoSeven®

- Dose: 90 $\mu\text{g}/\text{kg}$ for inhibitor
 - Repeat every 3–6 h; 6 h FVII half-life
- \$0.83/ μg
 - For our 80-kg patient, one dose = \$6000
- Cannot monitor
 - Generalized coag evaluation using PTT
 - No risk of DIC



Hemophilia A, B, and C

- *Anatomic* bleeding caused by congenital single-factor deficiencies
- 85% FVIII deficiency (hemophilia A)
 - 1 in 10,000 male births
- 14% FIX deficiency (hemophilia B)
 - Christmas disease, 1 in 30,000 male births
- 1% FXI deficiency (hemophilia C)
 - Rosenthal syndrome: 50% in Ashkenazi Jews
- Rare autosomal recessive single factor deficiencies
 - Prothrombin, V, VII, X, XIII

Hemophilia A Symptoms

Spontaneous anatomic (soft-tissue) bleeds

- Bleeding at umbilical stump and circumcision
- Delayed bleeding triggered by injury
 - Joints, large muscles, body cavities, GI, soft tissue, tongue, kidney, testicles, brain, CNS
- Spontaneous bleeds, especially into joints
- Inflammation, hematomas, hemarthroses

Severe	Moderate	Mild
70%	15%	15%
<1% FVIII	1–5% FVIII	6–30% FVIII
Spontaneous bleeds	Bleed after minor trauma	Bleed after major trauma



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Hemarthroses

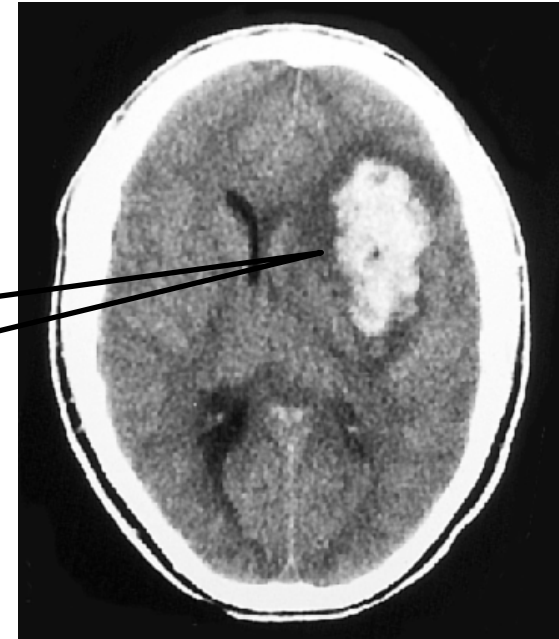


**Airway
Obstruction**

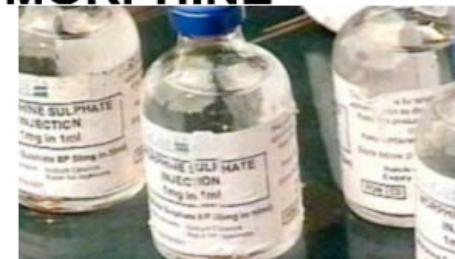
Cerebral Bleeds and Other Complications

- Lifestyle
- Economic
- Vocational
- Neurologic
- Psychological
- Lack of insurance
- Narcotics addiction

Cerebral
hemorrhage

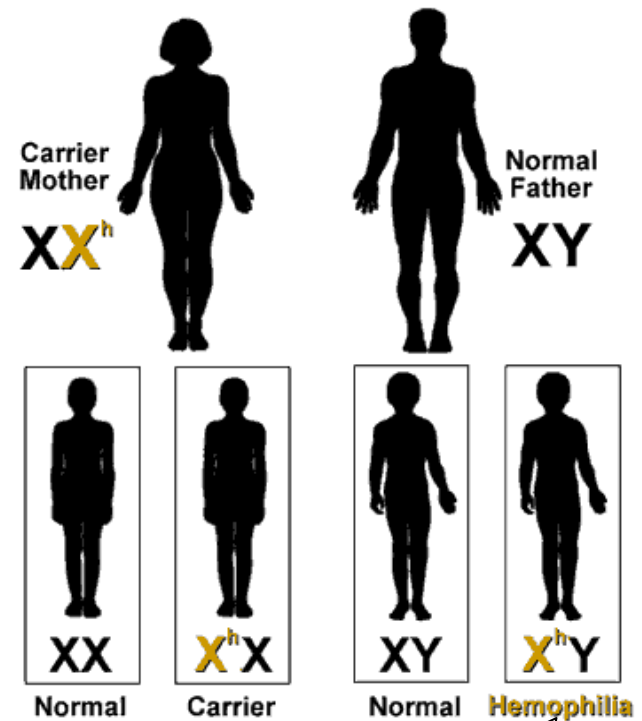


MORPHINE



Hemophilia A Inheritance

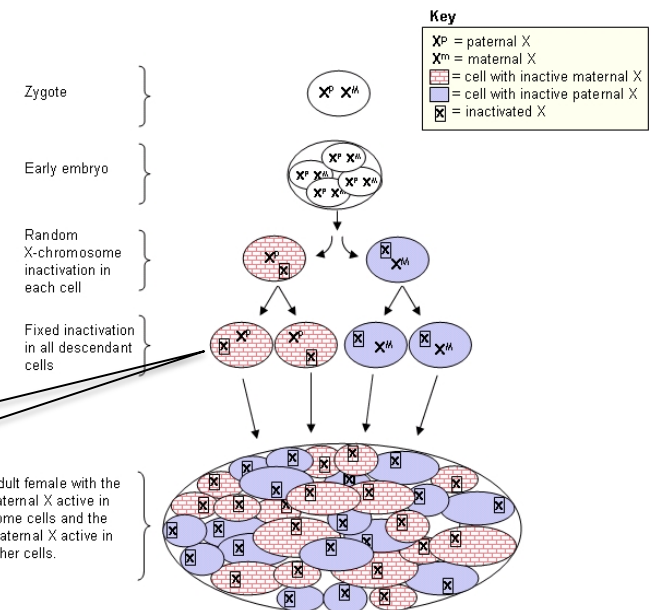
- Sex-linked recessive, 1/10,000 males
- Carrier mother, hemizygous son



Hemophilia A Genetics

- 186 kb gene on X chromosome
 - Deletions, stop codons; missense and nonsense point mutations
 - Predominantly quantitative deficiency, a few are qualitative
 - 25–30% *spontaneous mutation rate*
- Hemophilia A in females is rare
 - Acquired anti-FVIII inhibitor (IgG₄)
 - Autosomal hemophilia or VWD type N (Normandy)
 - Hemophilic father with carrier mother
 - Random “excess Lyonization”

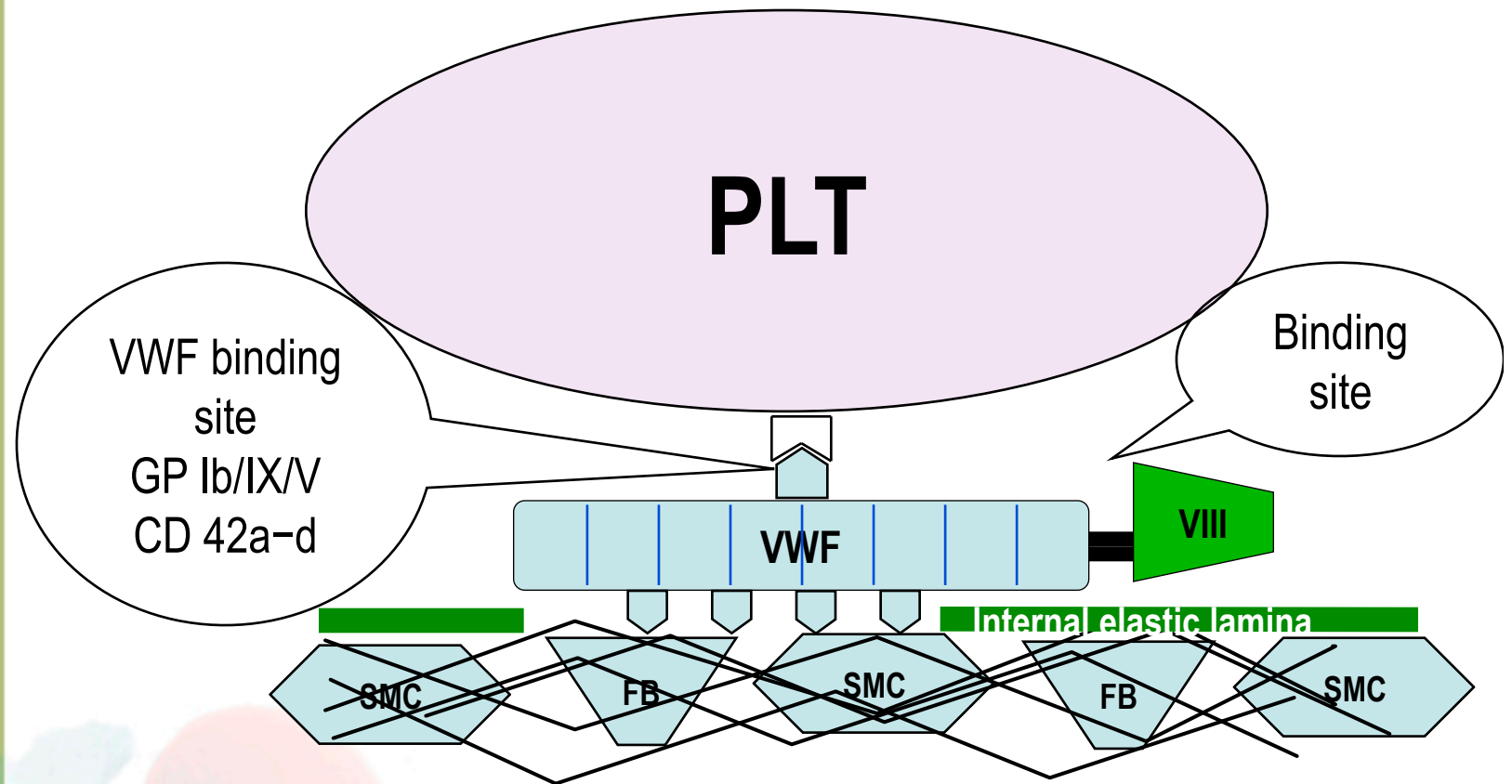
Unbalanced
excess of
maternal X



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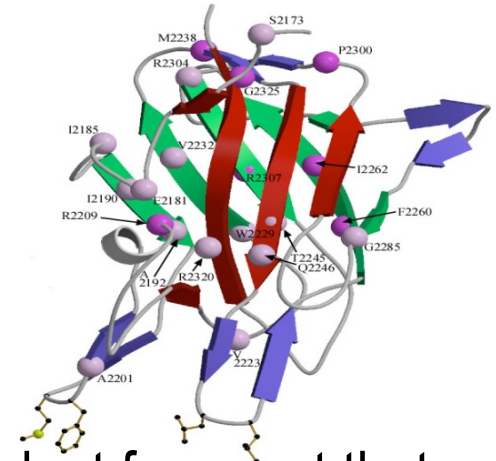
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FVIII Requires VWF Carrier



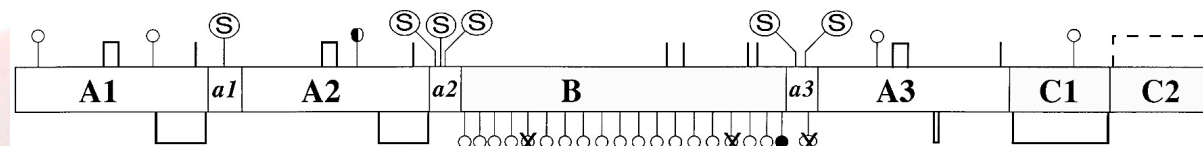
VWF: von Willebrand factor
SMC: smooth muscle cell
FB: fibroblast
Lnes: collagen

FVIII is a Glycoprotein Cofactor



- 285,000-D heterodimer
- Cleaved by thrombin, leaving a Ca⁺⁺-dependent fragment that detaches from VWF and binds factor IXa and platelet phospholipid: *phosphatidyl serine*
- Stabilizes IXa to form the “tenase” complex
- Deficiency slows thrombin production
- In vitro, deteriorates 5%/hour at 18–24°C

1 400 800 1600 2000 2332



glycosylated asparagine
 non-glycosylated
 disulfide bridge
 free cysteine
 partially glycosylated
 potentially glycosylated
 proposed disulfide bridge
 sulfated tyrosine

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Initiation: Exposed TF binds VIIa, activates IX→IXa and X→Xa

Extrinsic

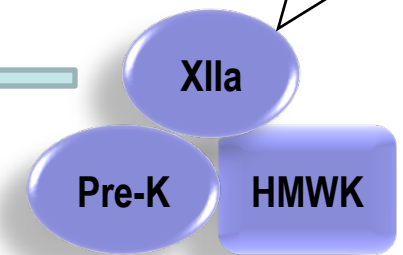


Common

Fritsma MG, Fritsma GA. Overview of hemostasis and coagulation. In Keohane EM, Smith LJ, Walenga JM. Rodak's Hematology, 5th Edition, Elsevier 2015

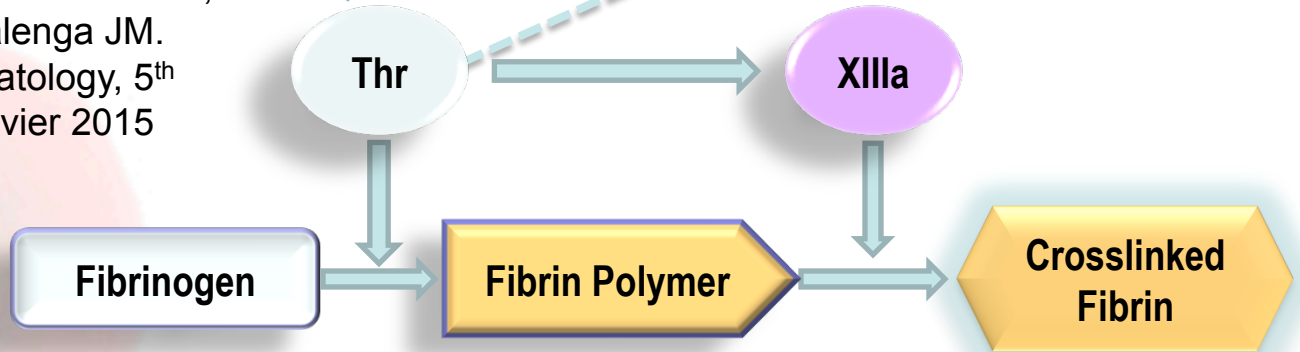
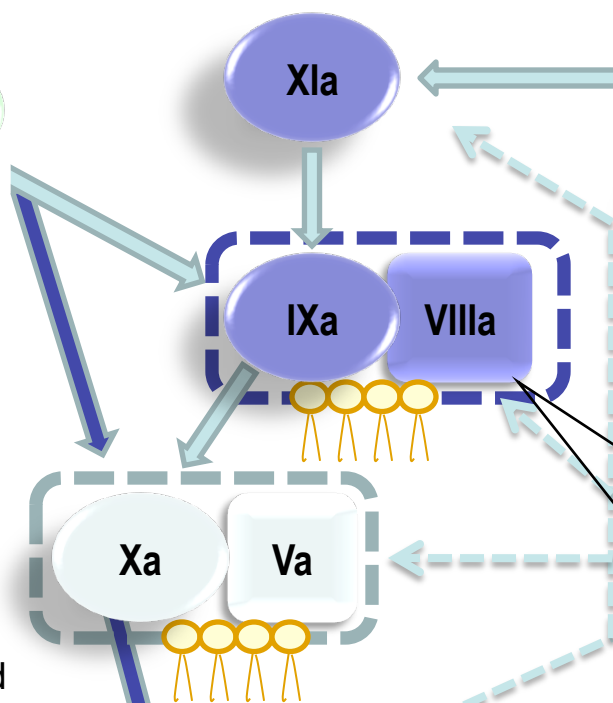
Coagulation

In vitro initiation: negatively charged particles



Intrinsic

VIIIa: thrombin-activated Ca⁺⁺-dependent heterodimer, detaches from VWF to bind PL & FIXa to form "tenase"



1st Documented Bleeder's Disease

2nd century: Talmudic ruling of *Rabbi Judah the Patriarch* exempts a 3rd son from circumcision if two elder brothers had died of bleeding after circumcision

2nd century: *Rabbi Simon ben Gamaliel* forbade a boy to be circumcised after sons of his mother's three elder sisters had bled to death after circumcision

11th century: Arabic surgeon *Albucasis* describes village males who bled to death from "trivial" wounds



Ingram GIC. The history of haemophilia. J Clin Pathol 1976; 29: 469-79.

1791–1803: British & American Families

- 1000–1800: Several written references to “bleeders”
- 1791 (Britain), Zoll: 6 brothers
 - Each bled to death after minor injuries
 - Half-siblings by a different mother were unaffected
- 1803 (Philadelphia), Otto: “A hemorrhagic disposition existing in certain families”
 - Recorded males in his own family with symptoms and recognized transmission through asymptomatic women
 - Traced pedigree to a woman named Smith in Plymouth, 1720–30
- 1820 (Germany) Nasse accurately defines the inheritance pattern, named “Nasse’s law.”

Otto quoted in Bulloch W, Fildes P. Treasury of human inheritance, parts V & VI, section XIVa, Haemophilia, 1911.

1800–28: Documented Names

- Haemorrhoea
- Bleeding disease
- Idiosyncrasia haemorrhagica
- Hereditary haemorrhagic diathesis
- 1828: first use of "haemophilia" (blood-loving) appears in the title of a treatise by Hopff at University of Zurich

Hopff F. Cited by United States Surgeon General's catalogue, 1st series: Hemophilia, 1828.

Bulloch and Fildes

Bulloch W, Fildes P. *Treasury of human inheritance, parts V and VI, section XIVa, haemophilia.*

Published as Eugenics Laboratory memoirs XII, Francis Galton Laboratory for National Eugenics, University of London; 1911, Dulau and Co, 37 Soho Square, London.

- 949 references & case reports from 235 pedigrees
- Identified hemophilia as hereditary and sex-linked
- Didn't understand hemophilia carrier status
- Meticulously traced the current spread of the mutation throughout Queen Victoria's family

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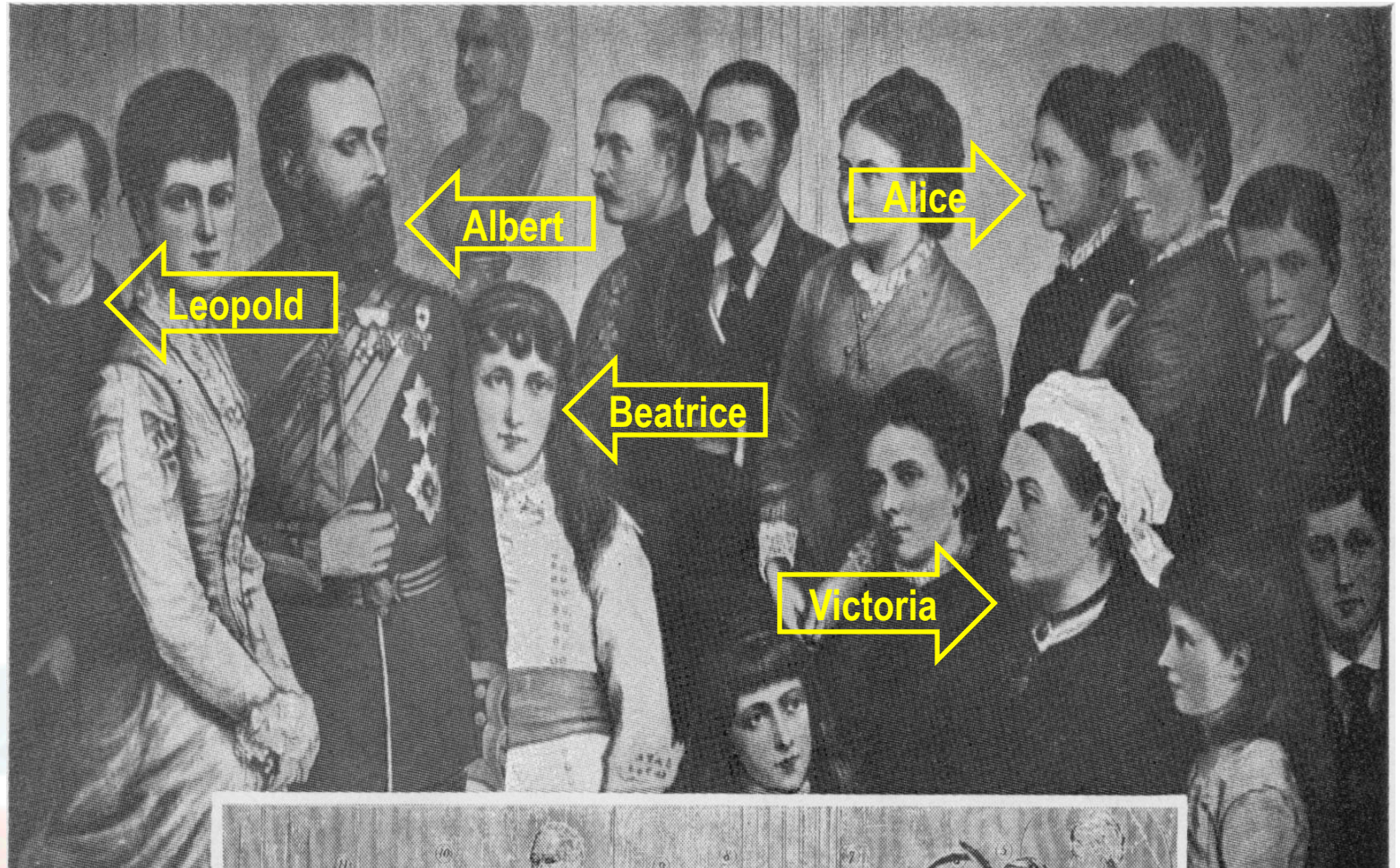
Alexandrina Victoria; May 24, 1819–Jan 22 1901, was *Queen of the United Kingdom of Great Britain and Ireland* from her coronation at 18, June 20, 1837 until her death, altogether 63 years and 7 months. The Victorian era was a time of UK industrial, political, imperial, and military progress.

Queen Victoria: Hemophilia Carrier

- Presumed spermatogenesis mutation in her father; Edward, Duke of Kent, in his 50s when Victoria was conceived
 - FVIII mutation in men double prevalence in women
- Victoria's seventh child, Leopold, was hemophilic
 - Stigmatized as a "weak" invalid by mother, hidden from public, married at 29 contrary to medical advice
 - Died of cerebral hemorrhage following a fall at 31
- Two daughters of Victoria, Alice (2nd) and Beatrice (8th) turned out to be carriers, as learned later

Massie RK. Nicholas and Alexandra. (1968). Gollancz, London.

Queen Victoria and Family



● Normal Female

● Carrier Female

Normal Male

Hemophiliac Male

Mutation?

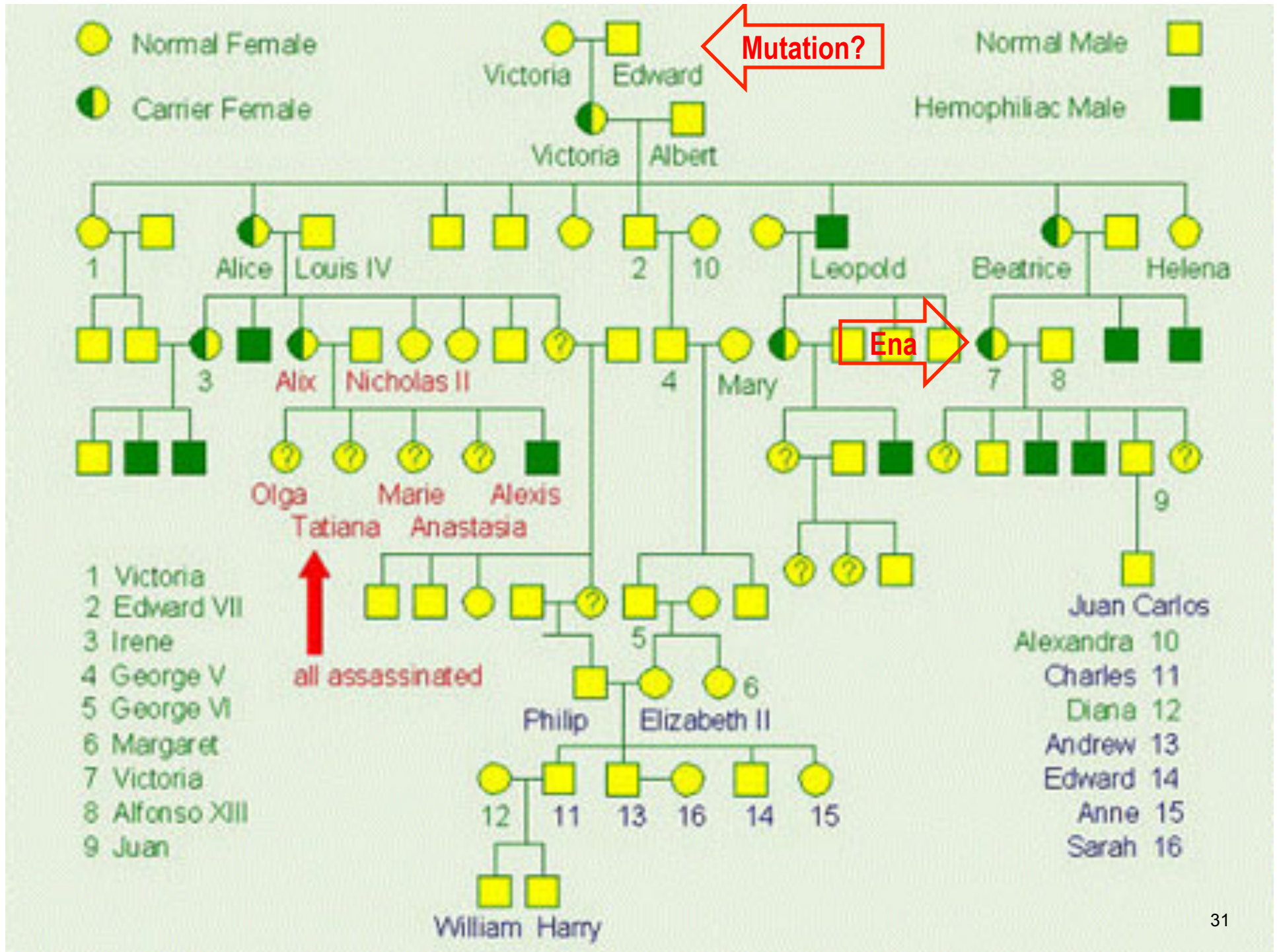
Ena

all assassinated

- 1 Victoria
- 2 Edward VII
- 3 Irene
- 4 George V
- 5 George VI
- 6 Margaret
- 7 Victoria
- 8 Alfonso XIII
- 9 Juan

- Juan Carlos
- Alexandra 10
- Charles 11
- Diana 12
- Andrew 13
- Edward 14
- Anne 15
- Sarah 16

William Harry



Victoria Eugenie (Ena) Battenberg

- Beatrice's daughter, 2 hemophilic brothers
- Alfonso XIII of Spain married her in 1906
 - Warned by Spanish Embassy, but trusted his luck
- Alfonso XIV b. 1907 bled from circumcision
 - Alfonso XIII blamed Ena, grew to despise her
 - More boys: Jaime, deaf-mute; Gonzalo, hemophilic
 - Spaniards blamed the British
- 1931, royal family in exile in Rome
 - Alfonso XIV was too ill to appear
 - 1933, marries Cuban commoner, abdicates
- Juan, born 1913, healthy
 - His son **Juan Carlos** was King of Spain 1975 'til abdication 6/2/2014



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The Romanovs: Tsar Nicholas II, Alexandra (Alix, granddaughter of Victoria) and Family



Tsarevich Alexis
b. 7/4/1904; 301
gun salute



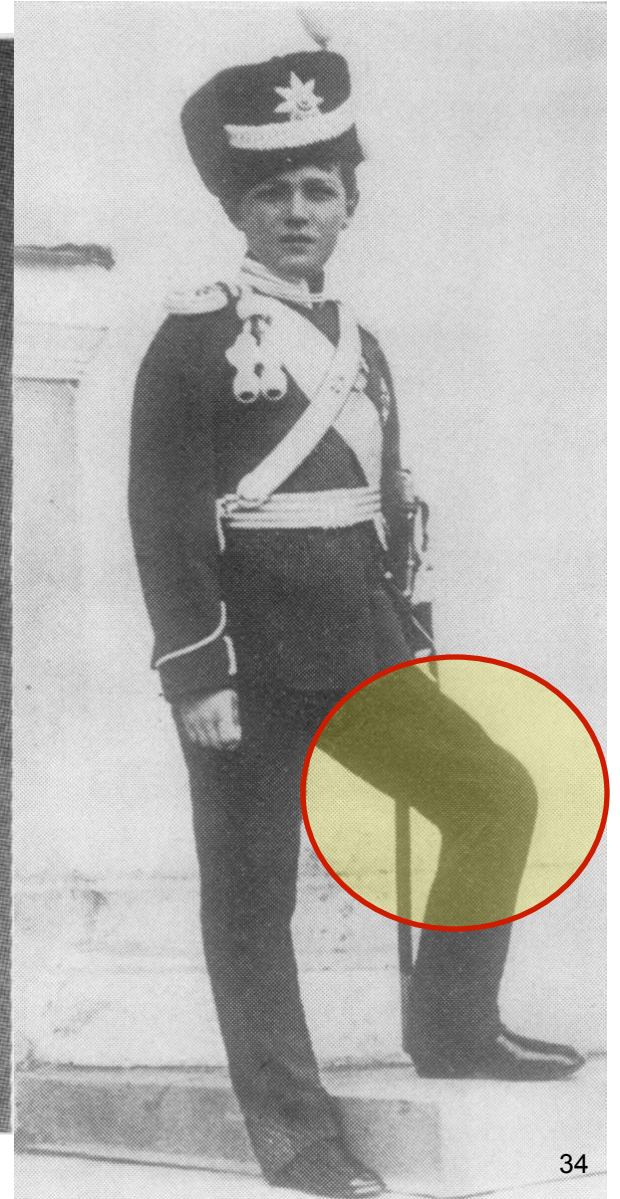
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Prince Alexis, 1912

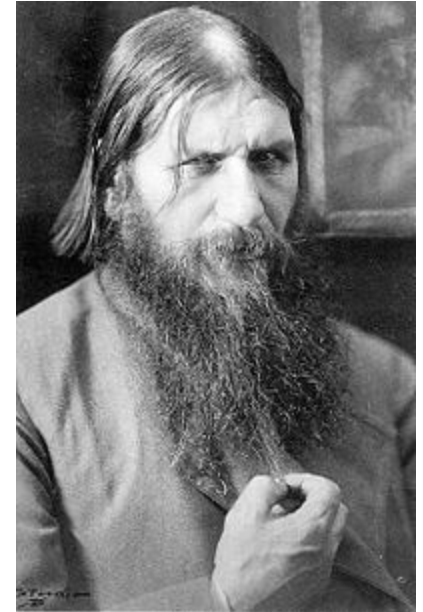


The Fritsma Factor



Grigori Yefimovich Rasputin 1869–1916

- 1869, Born Pokrovskoye, Siberia
 - Observed two sibs drown in nearby river
- 1887: 3 months in Verkhoturye monastery (in the Urals)
- 1901: *strannik* (pilgrim), wandered Greece, Jerusalem
- 1903: Saint Petersburg, *starets* (holy man) with healing & prophetic powers
- 1905: Introduced to Alexandra by close friend Anna Vyrubova to help with Alexis' hemophilia
- 1906–12, ever present to provide care

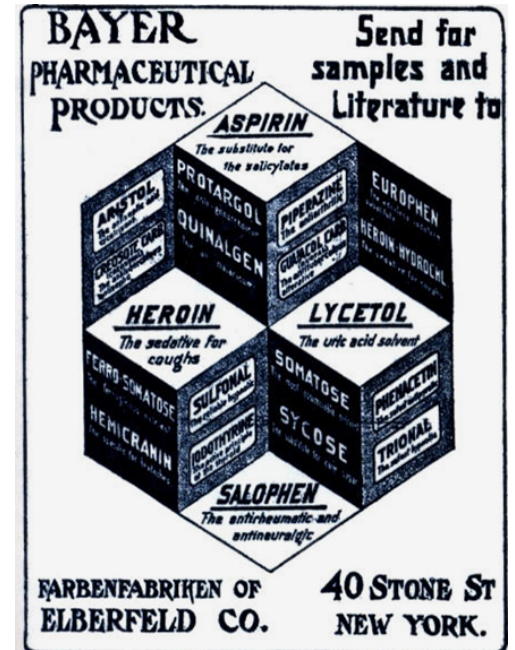


Rasputin's Power Rises

1912 Belovezhski Forest Holiday

- Alexis is near death, telegram to Rasputin in Siberia
- Calming influence, distraction?
 - “God has seen your tears and heard your prayers. Don't be sad, the little boy is not going to die. Don't let the doctors frighten him”
- Leeches?, Faith healer, hypnotism?

Rasputin became the czar's primary adviser and political gatekeeper. He engaged in monumental debauchery, and was hated by the Russian nobles, though loved as a religious figure by the Tsarina and many of the peasants.



The Romanovs 1912–16

Pierre Gilliard, Alexis' tutor, wrote in 1921: *“The illness of the Tsarevich cast its shadow over the whole of the concluding period of Tsar Nicholas II's reign. Without appearing to be, it was one of the main causes of his fall, for it made possible the phenomenon of Rasputin and resulted in the fatal isolation of the sovereigns who lived in a world apart, wholly absorbed in a tragic anxiety that had to be concealed from all eyes.”*

Historians have since disputed the contribution of Alexis' hemophilia to Russian politics, but the strain on the royal household is clear enough.



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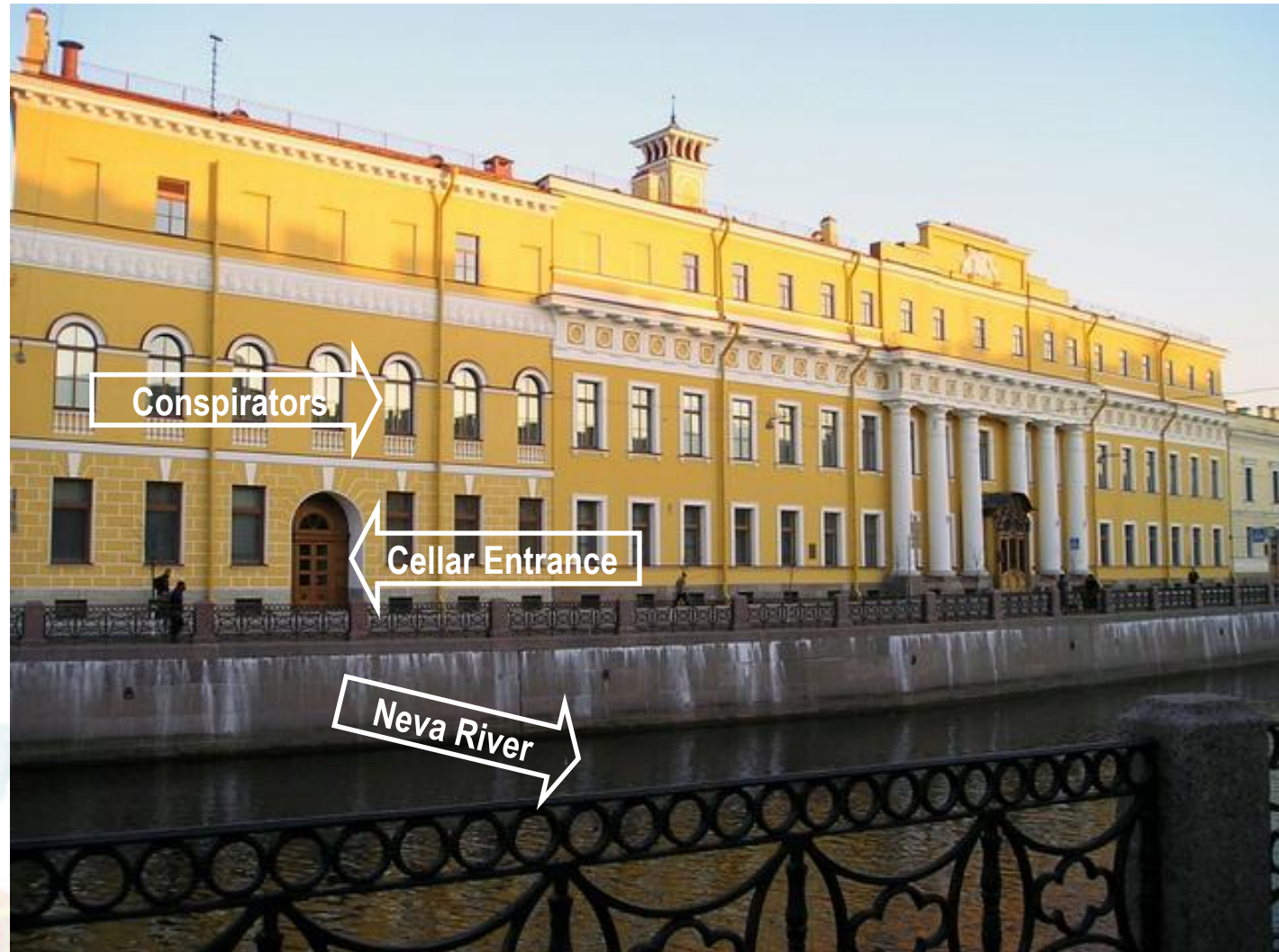
Rasputin and Admirers, 1914



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Yusupov Palace, St. Petersburg



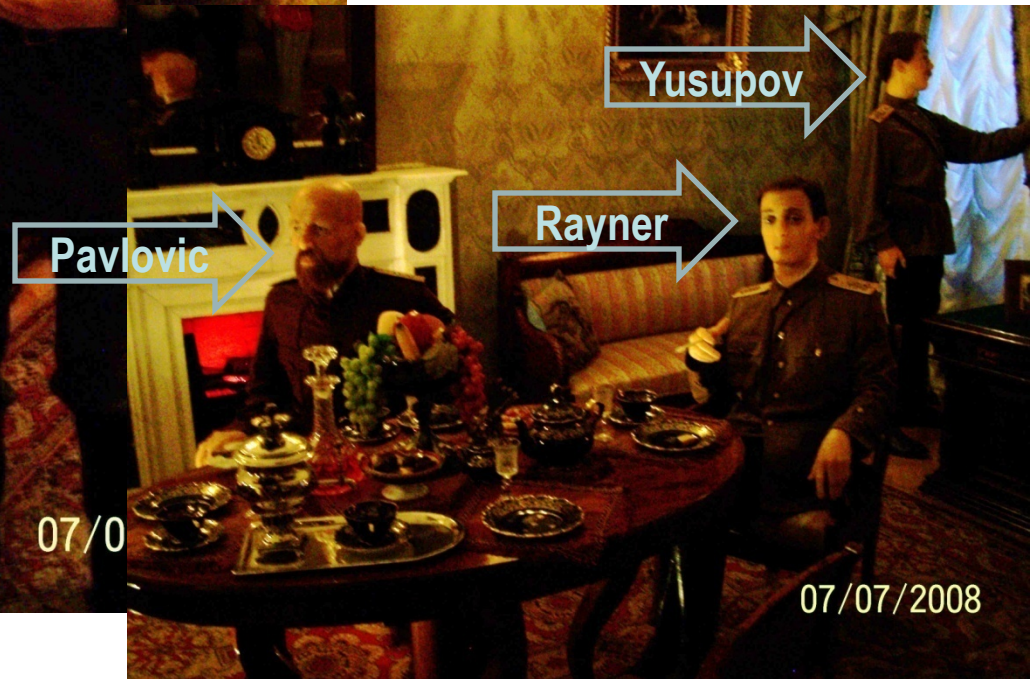
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Maj. Felix Yusupov: Dec 16, 1916



Wax figures at the Yusupov palace in St. Petersburg, photographed in 2008



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Poisoning Rasputin



Cyanide-laced cakes



Treatment Attempts 1901–1942

- Lime
- Gelatin
- Oxygen
- Splenectomy
- Bone marrow
- Sodium citrate
- Calcium lactate
- Witte's peptone
- Hydrogen peroxide
- Induced anaphylaxis
- Antidiphtheric serum
- The 'galvanic needle'
- Animal and human sera
- Adrenaline
- Bird's muscle
- IV oxalic acid
- Vitamin therapy
- X-ray irradiation
- Serum from the mother
- Tissue fibrinogen by mouth
- Bromide extract of egg white; sedative
- Blood—both withdrawn and injected therapeutically
 - “Auto-hemotherapy”
- Female hormone therapy
 - in the belief that femininity prevents expression of the hemophilic gene

Effective Hemophilia Treatments

- 1926, Surgeon General: 12 published attempts at whole blood transfusion beginning after WW I
 - Blood groups defined by Landsteiner in 1900
- 1934, McFarlane: topical Russell viper venom
- 1938, Patek and Taylor, Brinkhouse: first characterization of anti-hemophilic globulin
- 1938, McFarlane: fresh whole plasma
- 1944: EJ Cohn fractionation of plasma
 - Animasl: Biggs and Macfarlane, 1954; Bidwell, 1955
 - Kekwick and Wolf, 1957; Soulier, Gobbi, Larrieu, 1957; Blomback, Blomback, Nilsson, 1958

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1952: Stephen Christmas, Canadian Hemophilia Soc: Delineated FIX def (*Christmas disease*) from FVIII def (*Alexis was factor IX deficient*)



1953
Nine-year-old Donald Burris smiles on father's knee. His is believed to be first successful appendectomy operation to a haemophiliac in North America.

Breakthroughs: CRYO, Anti-hemophilic Factor

- 1964, Judith G. Pool (1919–75, U of Chi)
 - Developed cryoprecipitate
 - First opportunity for hemophilic home care
- 1968, Kenneth M. Brinkhous
 - 1908–2000, UNC Chapel Hill
 - First to chemically characterize factor VIII in 1938
 - Developed AHF with hemophilic dog experiments
 - AHF released through Hyland in 1968



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Advances in the 1960–70s

- 1960: prevent bleeds during dental extractions and minor procedures using Kohn fraction
- 1960: surgical orthopedic correction of hemarthroses using AHF
- 1970: home therapy
- 1973: hemophilia treatment centers
- 1980: prothrombin complex concentrate
 - Proplex[®], II, VII, IX, X for IX deficiency
- 1980: activated PCC, FEIBA[®] for inhibitors
- 1980: life expectancy was 60
 - But high hepatitis rate, 20,000 donors/pool
 - No viral inactivation

Rev. Robert K. Massie III



Robert K. Massie



Massie R, Massie K. *Journey*. Knopf, USA 1973

The Fritsma Factor



Susan Massie

HIV; AIDS and the CDC

- Fall 1980: *Pneumocystis carinii* pneumonia (PCP) and Kaposi sarcoma (KS) in homosexual males
 - Searched for non-infectious immunodeficiency causes such as amyl nitrite “poppers,” anti-sperm antibodies or anal intercourse
- Spring 1982: CDC recorded three cases of PCP in hemophilic men receiving FVIII concentrate, all died
 - Paralleled reports of similar symptoms in Haitian hemophiliacs and in US drug abusers
 - No homosexual behavior or illegal drug use
 - Led to concept of blood-borne viral infection

Evatt BL. The tragic history of AIDS in the hemophilia population, 1982–1984. *J Thrombos Haemost* 2006; 4: 2295–301.



CDC Report Delivered 7/27/82



- CDC reported to blood and plasma industries, gay organizers, hemophilia organizations, FDA and NIH; coined the term AIDS
- Consensus to defer action
 - Evidence of three patients too weak to conclude hemophiliacs were a risk group
 - Concern it would stigmatize homosexual community
 - Loss of homosexual donors to blood and plasma services
 - It would cost millions to change manufacturing practices
- Dec 1982: 6 more CDC cases: two non-hemophilic blood recipients, one an infant
 - Donor services refused to reveal donor identities to avoid embarrassing and subsequently losing donors
 - One definite identification slipped through, presumed infection

CDC Report #2, Delivered 1/4/83

- CDC reported the statistical prevalence of hepatitis B was the same in hemophilics and AIDS risk groups (surrogate association)
- CDC reported to the same groups including ARC, AABB, National Hemophilia Foundation, National Gay Task Force, Pharmaceutical Mfrs Association, Council of Community Blood Centers, State and Territorial Epidemiologists, and individuals.
- Again, consensus to not act, debate was irrational, acrimonious and public, harshly critical of Dr. Evatt and CDC

January 13, 1983

- CCBC and AABB: *“transfusions are life-saving procedures; some adverse reactions are acceptable to save lives. The rare disorder affecting nine cases is not enough to force a policy change.”*
- ARC head Dr Cumming wrote: *“It has long been noted that CDC increasingly needs a major epidemic to justify its existence... In short, we can not depend on the CDC to provide scientific, objective, unbiased leadership.”*
- The National Hemophilia Foundation, however, alarmed, had already contacted plasma processors in December, 1982



**American
Red Cross**



NATIONAL HEMOPHILIA FOUNDATION
for all bleeding and clotting disorders

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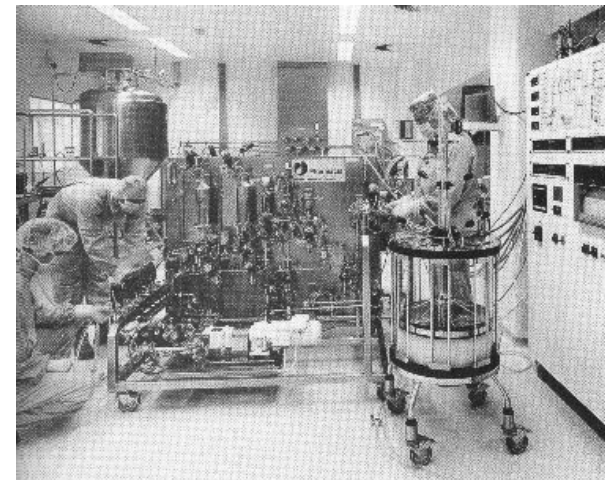
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National Hemophilia Foundation Initiative: 1983

- Dec, 1982: *Alpha Therapeutics* began to screen donors
 - Verbal, but 20% of commercial plasma came from donor services who refused to screen donors for sexual orientation
- US Public Health Service guidelines, March 4, 1983
 - CDC bypassed FDA, sent guidelines direct to PHS
 - Donor questionnaire and surrogate testing: hepatitis markers
- March, 1983, Baxter Hyland began heat treating plasma
- August, 1983, 26 confirmed cases of transfusion transmitted AIDS, including one FIX deficiency



The Fritsma Factor

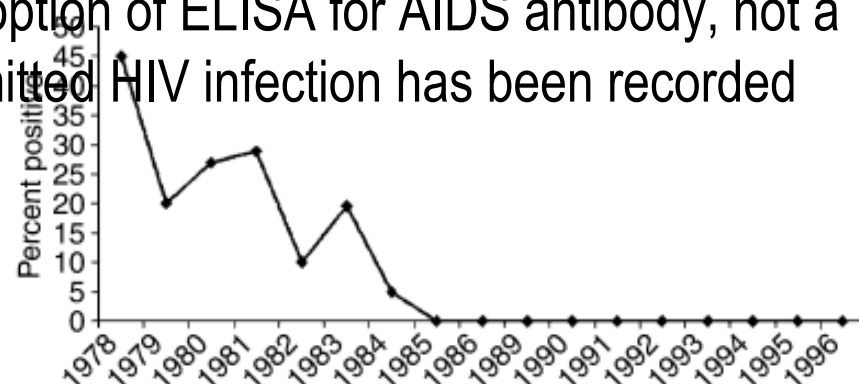


Virus Isolation: 1983

- July, 1983: Prof. Luc Montagnier of the Pasteur Institute isolates virus from lymphadenopathy patients
- Feb, 1984: Montagnier isolates the same virus from AIDS samples provided by CDC, confirmed by Robert Gallo, NIH
- Sep, 1984: Alpha and Cutter demonstrate heat treatment is safe and does not raise immunogenicity
- Oct, 1984: CDC/PHS screening and heat treatment guidelines are published and adopted by all agencies
- By 1984, 63% of 15,500 US hemophilia patients had HIV
- Since 1/1/1985, with adoption of ELISA for AIDS antibody, not a single new factor-transmitted HIV infection has been recorded



The Fritsma Factor



Frequency of HIV infection in US hemophilia birth cohorts. (From medical records)

Recombinant Clotting Factors

- 4/7/1976: *Genentech* incorporated
- 1981: *Genetics Institute* incorporated
- 1982: rFIX cloned by both (small molecule)
- Aug, 1984: Both cloned rFVIII gene and produced the protein, but yields are small
- 1985: rVWF coexpressed, improves rFVIII yields



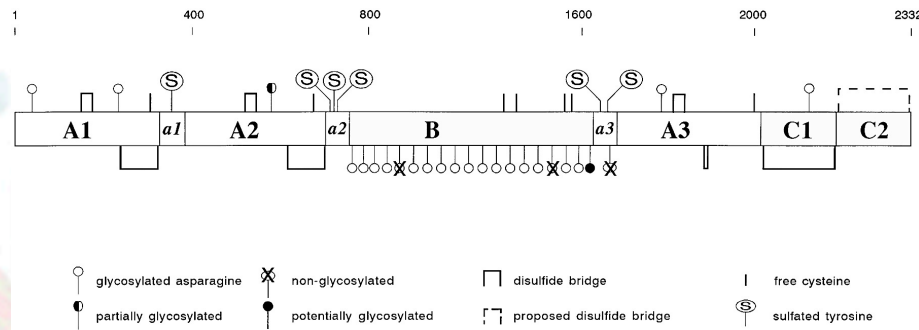
Pipe SW. Recombinant clotting factors. *Thromb Haemost* 2008; 99: 840–50.

rFVIII Barriers

- Extensive post-translational modification
 - Requires mammalian cloning system; Chinese hamster ovary cells
- rFVIII affinity for CHO membrane phospholipid
 - CHO membrane binds and degrades FVIII
 - CHO cells transfected with FVIII and VWF genes
 - Resolved by rVWF coexpression—higher affinity than membranes
- Need for serum-free cloning system
 - Eliminate animal viruses, ease purification
- High-volume fermentation process
 - Immunoaffinity, ion-exchange, Pasteurization, solvent-detergent, nanofiltration reduces 7000 L to 1 L.

rFVIII to Market

- Regulatory uncertainty
 - Sterility, consistency questions generate 600 tests/lot
- Baxter Hyland and Genetics Institute produced the first rFVIII, Recombinate
- Bayer Miles Lab/Cutter Biological and Genentech produced Kogenate
- 3/27/1987: UNC Chapel Hill, first phase II infusion into a 39 year-old hemophilic
- Recombinate approved 1992, Kogenate 1993



rFVIII Market Efforts and Inhibitor Formation

- B-domain deleted: Genetics Institute ReFacto, 2000
- No human or animal albumin in formulation: Advate, 2003
- Stabilizing matrix of sucrose, glycine, histidine: Kogenate FS
- Non-protein B-domain deleted: Wyeth Xyntha, 2008
- Hemostatic efficacy: 90% cessation of bleeding
- Risk of inhibitor formation in PUPs is approximately double the risk from plasma-derived FVIII concentrate
- Risk of additional inhibitor formation in response to rFVIII in previously treated persons (PTPs) is <1%

Pipe SW. The promise and challenges of bioengineered recombinant clotting factors. *J Thromb Haemost* 2005; 3: 1692–1701.

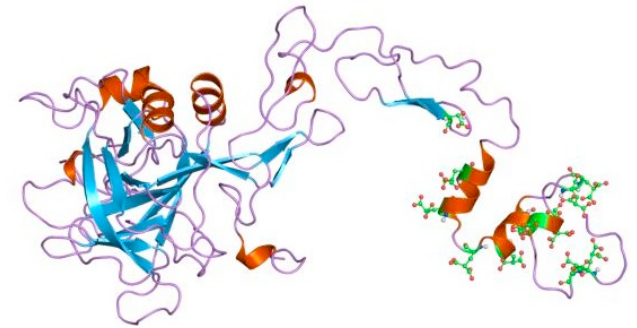
Barnes C, Lillicrap D, Pazmino-Canizares J, et al: Pharmacokinetics of recombinant factor VIII (Kogenate-FS®) in children and causes of inter-patient pharmacokinetic variability. *Haemophilia* 2006;2 (Suppl 4): 40–9.

THE FRITSMAN FACTOR

Your Interactive Hemostasis Resource



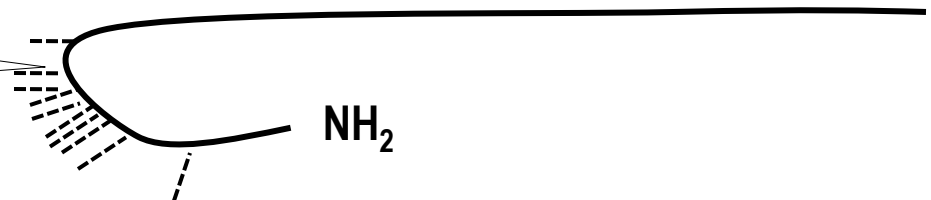
rFIX



- Genetics Institute BeneFix 1998
- Barrier: γ -carboxylation of 12–20 glutamic acids at amino terminus requires...
 - Vitamin K
 - Coexpressed γ -glutamyl carboxylase
 - Coexpression of furin activation enzyme
- 90% bleeding cessation: 1–2 infusions on demand
- PUPs—3% incidence of high titer inhibitors

12–20 GLA molecules

The Fritsma Factor



Inhibitor Formation in Severe Hemophilia

- Meta-analysis of 20 trials; pd versus rFVIII
- 1248 pts, 798 severe, median age 12
- 632 treated w/ pdFVIII and 616 w/ rFVIII
- 14% inhibitors in pdFVIII arm, 25% in rFVIII
- High titer: 8.8% in pdFVIII, 12.3% in rFVIII

lorio A, Marcucci M, Marchesini E, Mannucci P. Plasma derived and recombinant factor VIII concentrates include a different rate of inhibitor development in hemophilia A patients. A systematic review of the literature. J Thromb Haemost 2009;7

Primary Prophylaxis in Children

- Abundant rFVIII/IX; concern for VAD clotting
- 2005: Joint damage outcome study
 - 25 IU/kg every other day generates 6X decrease in joint deterioration by MRI vs on-demand (OD) Rx up to 6 YO
- 2009 Italian study on prophylaxis
 - 10 Y f/u on 25 IU/kg 3X a week vs OD: 0.52 vs 1.08 total bleeds and 0.2 vs 0.52 joint bleeds/patients/month
- 2009 Danish/Russian study
 - Orthopedic issues 15.6 in OD vs 2.2 in prophylaxis

Franchini M, Coppola A, Molinari AC, et al. Forum on the role of recombinant factor VIII in children with severe haemophilia A. *Haemophilia* 2009; 1–9.

Gringeri A, Lundin V, von Mackensen S, et al. Primary and secondary prophylaxis in children with haemophilia A reduces bleeding frequency and arthropathy development compared to on demand treatment; a 10-year, randomized clinical trial. *J Thromb Haemost* 2009;7

Ingerslev J, Lethagen S, Poulsen L, et al. A case-controlled Danish-Russian comparative study of clinical outcomes in younger severe haemophilia patients treated with prophylaxis compared to those managed with on-demand treatment. *J Thromb Haemost* 2009;7

On-demand Versus Primary in Adults

- 19 hemophilics 30–45 YO, all <1 unit/dL
 - Mean 2 bleeds/month, no additional bleeding disorder
 - ≥3 bleeds in elbow, ankle or knee, prior 6 months
 - Bayer Kogenate FS: sucrose-stabilized rFVIII
 - Prospective, open-label
- Six months OD Rx per bleed
 - 20–100 units/kg every 12–24 h to cessation
- Six months' prophylaxis
 - Home: 20–40 units/kg 2 mL/min infusion 3x/week
 - 1st month of prophylaxis was run-in, six months data collection
- Outcomes
 - Primary: patient self-assessed joint bleeds
 - Secondary: all bleeds, joint function, QOL, health economics, safety

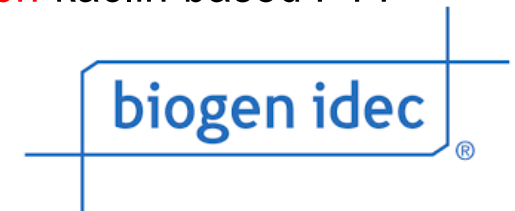


Results

Median of 19 subjects	6 m on-demand	6 m prophylaxis
Infusions	29	88
Total consumption, units	70,421	211,933
Joint bleeds	15	0
All bleeds	20.5	0
Spontaneous bleeds	13.5	0
Trauma bleeds	2.5	0
<ul style="list-style-type: none"> • Baseline: median joint bleeds: 14.0; 24% mild, 63% moderate, 13% severe • Fourteen used rFVIII, two used plasma-derived FVIII, three used both • Prophylaxis median trough: 48 h: 6 units, 72 h: 4 units • Mean total Gilbert joint function score (pain, swelling, atrophy, deformity, range of motion, instability): on-demand, 25.3%, prophylaxis 19.8% • Safety: no Rx-related adverse events 		

Extended Half-life Factors VIII and IX

- Recombinant B-domain deleted Fc fusion factor VIII
 - Biogen Idec Eloctate, FDA-approved 2014
 - 96 adult hemophilic males with > 12 annual bleeds, 3–4 doses/week
 - Extended by Fc receptor and IgG recycling pathway
 - rFVIII-Fc half-life 19 h versus rFVIII 12; 1.6–3.6 annual bleeds
 - Prophylactic Rx frequency up to 5 days versus 3–4 doses/week
 - Monitor using clot-based FVIII assay with **non**-kaolin-based PTT
- Recombinant Fc fusion factor IX
 - Biogen Idec Alprolix, FDA-approved 2014
 - Monitor using FIX assay with **non**-kaolin-based PTT or chromogenic
 - rFIX-Fc half-life 70–80 h versus 24 h, 7–10 day intervals



Shapiro AD¹, Ragni MV, Kulkarni R, et al. Recombinant factor VIII Fc fusion protein: extended-interval dosing maintains low bleeding rates and correlates with von Willebrand factor levels. *J Thromb Haemost.* 2014;12:1788–800.

Mancuso ME, Mannucci PM. Fc-fusion technology and recombinant FVIII and FIX in the management of the hemophilias. *Drug Des Devel Ther.* 2014 28;365–71.

Gene-based Hemophilia Therapy

- Genetic delivery targets: liver, skeletal muscle, hematopoietic tissue, endothelial cells
- Vectors: adeno-associated virus (AAV) and liver sinusoidal endothelial cells (LSEC)
- Barriers: theoretical T-cell immune response to vector, FVIII molecule too large for vector, inefficient transduction
 - Fomin ME, Togarriti P, Muench MO. Progress and challenges in the development of a cell-based therapy for hemophilia A. *J Thromb Haemost* 2014; 12: 1954–64.
 - Chuah MK, Nair N, Vandendriessche T. Recent progress in gene therapy for hemophilia. *Hum Gene Ther* 2012; 23:557–65.
 - Lozier J. Gene therapy of the hemophilias. *Semin Hematol* 2004; 41:287-96.

FIX Gene Transfer: Landmark Study

- Adenovirus-associated virus vector (AAV)
 - Codon-optimized human factor IX transgene
 - scAAV2/8-LP1-hFIXco
 - FVIII gene is 7 kb, AAV capacity 4.6 KB
- Six hemophilia B (<1% FIX) patients 29–64
 - 2 with FIX null, 4 with missense mutations (type II deficiency)
 - All receiving prophylactic FIX 3X/week
 - No inhibitors, no antibodies to AAV
- Vector administration dosages
 - 2×10^{11} vector genomes/kg body weight (vg/kg)
 - 6×10^{11} vg/kg
 - 2×10^{12} vg/kg

FIX Transfer Study Outcomes

- 4 of 6 required no FIX Rx
- 2 required on-demand FIX Rx
 - One ran the half-marathon
- High-dose subjects had rise in ALT
 - Immune response to hepatocytes?
 - Controlled with prednisolone
- No anti-FIX antibodies
- Did not detect cellular immunity to vector

Nathwani, AC, Tuddenham GD, Rangarajan S, et al, Adenovirus-associated virus vector-mediated gene transfer in hemophilia B. NEJM 2011;365: 2357-65.

BioMarin BMN270 FVIII Transfer Trial

- B-domain-reduced FVIII with minimal glycosylation
- Codon optimization: AAV8 variant adequate capacity
 - University College London and St. Jude's Research Hospital
 - Successful in mice and macaques
- Phase 1 & 2 study: enrolled first severe hemophilia A patient 9/28/15, inviting 11 more



McIntosh J, Lenting PJ, Rosales C, et al. Therapeutic levels of FVIII following a single peripheral vein administration of rAAV vector encoding a novel human factor VIII variant. *Blood*. 2013 25;121:3335-44

Ward NJ, Buckley SM, Waddington SN, et al. Codon optimization of human factor VIII cDNAs leads to high-level expression. *Blood* 2011;117:798–807.

Additional rFVIII Preparations

Name	MFR	Comments: all rFVIII except Bax 111	Progress
Bax 111	Baxter	rVWF (not rFVIII)	At FDA
Bay 81	Bayer	Full-length rFVIII w/o human and animal proteins, Reduced inhibitors, normal to slightly extended half-life	At FDA
NovoEight	Novo Nordisk		Released
NuWiq	Octapharma		Released
Turoctocog	Novo Nordisk	Glycopegylated ;18.4 h half-life, ~1.5 x current Rx, reduced inhibitors	At FDA
Bay 94	Bayer	Pegylated, plasma/albumin free, full-length rFVIII, up to 7.5 d frequency	At FDA
Bax 855	Baxter	Pegylated plasma/albumin free, full-length rFVIII, 1.5 X Advate half-life	At FDA
rFVIII single-chain	CSL Behring	rFVIII covalent bond to VWF reduces clearance, extends half-life; no inhibitors	Phase II
ACE 910	Genentech	Novel recombinant protein mimics FVIII, activates IX and X, bypasses inhibitors, weekly SC injections	FDA Breakthrough