



THE FRITSMA FACTOR
Your Interactive Hemostasis Resource

Managing Hemophilia Therapy Rasputin to Recombinants

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The Fritsma Factor; Your Interactive Hemostasis Resource
www.fritsmafactor.com

Hemophilia case study
Hemophilia in history
Alexis and Rasputin
AIDs and recombinants
Extended half-life recombinants
Gene transfer

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40 YO Af-Am Hemophilic

- Bleeding into ankle: midnight
 - Anatomic soft-tissue and joint bleeds
 - Tried RICE, DDAVP (Stimate®) inhaler
- Ran out of factor VIII (FVIII) concentrate
 - Provided free by Medicare through hemophilia centers
 - National Hemophilia Foundation: www.hemophilia.org
- Clinical path resident on-call; night tech
 - Determine patient factor VIII activity
 - Order antihemophilic factor (AHF, FVIII) concentrate
 - Dispensed by transfusion service
 - Compute and prepare dosage
 - Reconstitute with sterile water, administer as IV push
 - Subsequently determine therapeutic factor VIII activity

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Hemophilia A Symptoms

Spontaneous anatomic (soft-tissue) bleeds




- Bleeding at umbilical stump, circumcision
- Delayed bleeding following injury
 - Joints, muscles, body cavities, GI, soft tissue, tongue, kidney, testicles, CNS
- Spontaneous bleeds, especially joints
- Inflammation, hematomas, hemarthroses

| Mild | Mod | Severe |
|--------------------------|--------------------------|--------------------|
| 6–30% VIII | 1–5% VIII | <1% VIII |
| Bleed after major trauma | Bleed after minor trauma | Spontaneous bleeds |
| 15% | 15% | 70% |



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Hemarthroses


Airway Obstruction

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Other Complications

- Lifestyle
- Economic
- Vocational
- Neurologic
- Psychological
- Lack of insurance
- Narcotics addiction



Cerebral hemorrhage



MORPHINE

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Calculating FVIII Concentrate Dosage

- One unit = activity in 1 mL normal plasma or 100%
- Determine plasma volume based on weight
 - Blood volume (BV, mL) = weight (kg) x 70 mL/kg
 - Use 60 mL/kg for obese, BMI 25–30
 - Use 50 mL/kg for morbidly obese, BMI >30
 - Plasma volume (PV, mL) = BV x (100%–HCT%)
- Determine units of FVIII concentrate required

Units of FVIII required =
(desired FVIII in units/mL – initial units/mL) x PV (mL)

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40 YO Af-Am Hemophilic Compute FVIII Dosage

- 80 kg, HCT 40%, factor level <1%
- Determine plasma volume
 - Blood volume (mL) = 80 kg x 70 mL/kg = 5600 mL
 - Plasma volume (mL) = blood volume (5600 mL) x (100%-40%) = 60% x 5600 = 3360 mL
- Determine units of FVIII required:
 - Wish to reach 80% factor level (0.8 units/mL), therefore...
 - Units of FVIII required = (0.8 units/mL - 0) x 3360 mL = 2688 (2700) units
 - Typical concentrate vial provides ~1000 units, use 3
- Avoid overdose: thrombotic and wasteful

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If Factor Assay not Available

- When factor assay is not available and timing is critical, assume 0 activity or...
- Estimate FVIII from PTT

| Factor VIII | PTT |
|-------------|-------|
| 40% | 35 s |
| 30% | 50 s |
| 20% | 65 s |
| 10% | 90 s |
| 1% | 120 s |

Example only, do not use
- Collect baseline plasma
 - Assay or freeze and confirm with assay next day shift
- Maintain patient database

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Plasma-derived FVIII Concentrates

- Theoretical risk of HBV, HCV, HIV transmission
 - Hemofil-M®, Monarc-M® (Baxter), Monoclade-P®
 - Human and animal plasma matrix
 - Purification: immunoaffinity column, solvent-detergent, Pasteurization, viral filtration, combinations
 - \$0.35-\$0.60/AHF unit
- Seroconversions per CDC surveillance: 0
 - Data from 2003
 - Predicted risk, 1:60,000

MONARC-M
[Antihemophilic Factor (Human)]
Method M, Monoclonal Purified

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Recombinant FVIII Concentrates

- Serum in culture medium
 - Helixate®, Kogenate®, Recombinate®
 - \$0.68-1.05/AHF unit: *some select for only previously untreated patients (PUPs)*
 - No HBV, HCV, HIV seroconversions
- No protein in culture or prep
 - Calculated viral risk=0, actual=0: Advate®
- B-domain-deleted FVIII concentrate
 - Human albumin: ReFacto
 - No protein: Xyntha
 - Extended half-life: Eloctate
 - Can't assay using clot-based FVIII assay, use chromogenic FVIII

Recombinate

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Factor VIII Assay

- Dilute plasma 1:10, add factor VIII-depleted reagent plasma 1:1
- Add PTT reagent, incubate 3 minutes
- Add CaCl₂, record interval to clot formation
- Compare result in seconds to calibrator curve

Factor VIII Activity Reference Curve

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Factor VIII Assay Plasma Dilutions Parallelism Indicates No Inhibitor

| Plasma Dilution | Seconds | Raw Factor VIII Activity | Computed Factor VIII Activity (x dilution) |
|------------------|---------|--------------------------|--------------------------------------------|
| 1:10 "undiluted" | 90 s | 20% | 20% |
| 1:20 | 104 s | 10% | 20% (parallel)* |
| 1:40 | 107 s | 5% | 20% (parallel) |
| 1:80 | 110 s | 2.5% | 20% (parallel) |

* <10% difference from undiluted indicates parallelism, no inhibitor

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40 YO Af-Am FVIII Concentrate Therapy

- Peak: 15 m after administration: 0.3 units/mL
 - Should have been 0.8 units/mL, what happened?
 - Suspect anti-factor VIII inhibitor
 - If peak reaches expected value, go to next administration
- Nadir (trough): 12 h after administration
 - Reflects half-life, should reach 50% of desired activity
 - Administer new AHF, use half the dosage second time

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FVIII Assay Plasma Dilutions non-Parallelism Indicates Inhibitor

| Plasma Dilution | Seconds | Raw Factor VIII Activity | Computed Factor VIII Activity (x dilution)* |
|------------------|---------|--------------------------|---------------------------------------------|
| 1:10 (undiluted) | 80 s | 10% | 10% |
| 1:20 | 93 s | 8% | 16% |
| 1:40 | 107 s | 5% | 20% |
| 1:80 | 108 s | 4% | 32% |

* >10% difference from undiluted = non-parallel & rising, implies inhibitor

Kasper CK. Laboratory diagnosis of factor VIII inhibitors. In Kessler C, Garvey MB, Green D, Kasper C, Lusher J. Acquired Hemophilia 2nd Edition. Excerpta Medica 1995

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40 YO Af-Am, FVIII Inhibitor

- Inhibitor: alloantibody to FVIII concentrate
 - 30% incidence, mostly severe hemophilia
 - FVIII dose response
- Factor VIII assay
 - If non-parallel, reflex to Bethesda titer
- The “poor man’s” Bethesda titer substitute
 - Dilute plasma 1:20 in normal plasma, perform PTT
 - If prolonged, presume >5 BU, treat as high-titer inhibitor
 - Rx with corticosteroids and FEIBA® or NovoSeven® (VIIa)
 - If PTT normal, presume <5 BU, treat as low-titer inhibitor
 - Rx with factor VIII concentrate
 - Confirm with full Bethesda titer
 - Larry D. Brace, PhD, Edward Hospital, Naperville, IL

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FVIII Inhibitor Therapy

- Plasma-derived factor IX complex
 - Activated prothrombin complex concentrate (PCC)
 - Prepared by BaSO₄ extraction
 - Available since 1980
 - FEIBA®, Autoplex®
- FEIBA dosage
 - 50 units/kg/12 h standard
 - 70 units/kg/8 h in hemorrhage
 - Limit 200 units/kg/24 h to avoid DIC risk: fatal
 - Cannot monitor: only general coag evaluation with PTT
 - \$0.78/unit, about \$4300/dose for the 80 kg patient

Ludlam DA, Morrison AE, Kessler C. Treatment of acquired hemophilia. Semin Hematol 1994;31 (Suppl 4) 16-19

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Recombinant FVIIa Concentrate

- Dose: 90 µg/kg for inhibitor
 - Repeat every 3-6 h
 - 6 h FVII half-life
- \$0.83/µg
 - For our 80-kg patient, one dose = \$6000
- Cannot monitor
 - General coag evaluation using PTT
 - No risk of DIC

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Hemophilia A, B, and C

- Anatomic bleeding caused by congenital single-factor deficiencies
- 85% factor VIII deficiency (hemophilia A)
 - 1 in 10,000 male births
- 14% factor IX deficiency (hemophilia B)
 - Christmas disease, 1 in 30,000 male births
- 1% XI deficiency (hemophilia C)
 - Rosenthal syndrome: 50% in Ashkenazi Jews
- Rare autosomal recessive single factor deficiencies
 - Prothrombin, V, VII, X, XIII

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Hemophilia A Inheritance

- Sex-linked recessive, 1/10,000 males
- Carrier mother, hemizygous son

Carrier Mother XX^* Normal Father XY

Normal XX Carrier XX^* Normal XY Hemizygote XY^*

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Hemophilia A Genetics

- 186 kb gene on X chromosome
 - Deletions, stop codons; missense and nonsense point mutations
 - 25-30% spontaneous mutation rate
 - Predominantly quantitative deficiency, a few qualitative
- Hemophilia A in females is rare
 - Acquired anti-FVIII inhibitor (IgG_4)
 - Autosomal hemophilia or VWD type N (Normandy)
 - Random "excess Lyonization"
 - Hemophilic father with carrier mother

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FVIII Requires VWF Carrier

PLT

VWF binding site
GP Ib/IX/V
CD 42a-d

VWF

FVIII

Subendothelial matrix

SMC: Smooth muscle cell
FB: Fibroblast
Lines: Collagen

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FVIII is a Glycoprotein Cofactor

- 285,000-D heterodimer
 - Translated from the X chromosome
- Cleaved by thrombin, leaving a Ca^{++} -dependent portion that detaches from VWF and binds factor IXa and phospholipid
- Stabilizes IXa in the "tenase" reaction
- Deficiency slows thrombin production
- In vitro, deteriorates 5%/hour at 18-24°C

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Coagulation

Initiation: Exposed TF binds VIIa, activates IX → IXa and X → Xa

In vitro initiation: negatively charged particles

Extrinsic

Common

Intrinsic

VIIa

TF

IXa

VIIIa

Xa

Va

Thrombin (IIa)

XIIa

XIa

XIIIa

XIVa

Fibrinogen

Fibrin Polymer

Crosslinked Fibrin

Fritsma MG, Fritsma GA. Overview of hemostasis and coagulation. In: Keoghane EM, Smith LJ, Walenga JM, Rodak's Hematology, 5th Edition, Elsevier 2015

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1st Documented Bleeder's Disease

2nd century: Talmudic ruling of *Rabbi Judah the Patriarch* exempts a 3rd son from circumcision if two elder brothers had died of bleeding after circumcision

2nd century: *Rabbi Simon ben Gamaliel* forbade a boy to be circumcised after sons of his mother's three elder sisters had died after circumcision

11th century: Arabic surgeon *Albucasis* describes village males who bled to death from "trivial" wounds

Ingram GIC. The history of haemophilia. J Clin Pathol 1970; 23: 403-13.

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1791–1803: British & American Families

- 1000–1800: Several written references to “bleeders”
- 1791 (Britain), Zoll: 6 brothers
 - Each bled to death after minor injuries
 - Half-siblings by a different mother were unaffected
- 1803 (Philadelphia), Otto: “A hemorrhagic disposition existing in certain families”
 - Recorded males in his own family with symptoms and recognized transmission through asymptomatic women
 - Traced pedigree to a woman named Smith in Plymouth, 1720–30

Otto quoted in Bulloch W, Fildes P. *Treasury of human inheritance*, parts V & VI, section XIVa, Haemophilia, 1911.

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1800–28: Documented Names

- Haemorrhoea
- Bleeding disease
- Idiosyncrasia haemorrhagica
- Hereditary haemorrhagic diathesis
- 1828: first use of “haemophilia” (blood-loving) appears in the title of a treatise by Hopff at University of Zurich

Hopff F. Cited by United States Surgeon General's catalogue, 1st series: Hemophilia, 1828.

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Bulloch and Fildes

Bulloch W, Fildes P. *Treasury of human inheritance*, parts V and VI, section XIVa, haemophilia. Published as Eugenics Laboratory memoirs XII, Francis Galton Laboratory for National Eugenics, University of London; 1911, Dulau and Co, 37 Soho Square, London.

- 1000 references and case reports
- 200 pedigrees
- Identified haemophilia as sex-linked, but carrier status not understood
- Meticulously traces the current spread of the mutation throughout Queen Victoria's family

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Alexandrina Victoria; May 24, 1819–Jan 22 1901, was *Queen of the United Kingdom of Great Britain and Ireland* from her coronation at the age of 18, June 20, 1837 until her death, altogether 63 years and 7 months. The Victorian era was a time of United Kingdom industrial, political, imperial, and military progress.

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Queen Victoria: Hemophilia Carrier

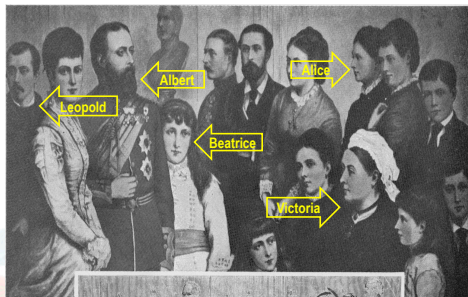
- Presumed spermatogenesis mutation in father; Edward, Duke of Kent, in his 50s when Victoria was conceived
- Victoria's seventh child, Leopold, was hemophilic
 - Stigmatized as a “weak” invalid by his mother
 - Married at 29
 - Died of cerebral hemorrhage following a fall at 31
- Two daughters of Victoria, Alice (2nd) and Beatrice (8th) turned out to be carriers

Massie RK. Nicholas and Alexandra. (1968). Gollancz, London.

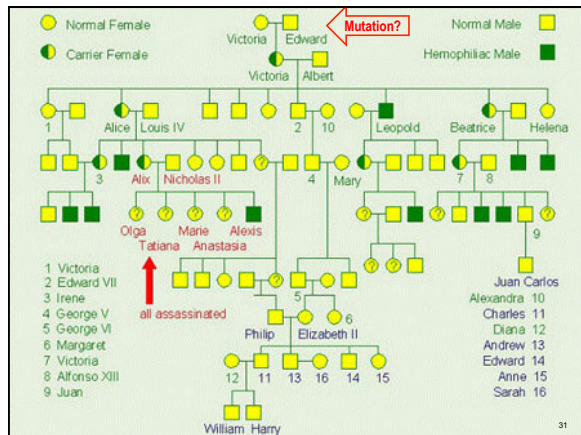
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Queen Victoria and Family




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Victoria Eugenie (Ena) Battenberg


- Beatrice's daughter, 2 hemophilic brothers
- Alfonso XIII of Spain married her in 1906
 - Warned by Spanish Embassy, but trusted his luck
- Alfonso XIV b. 1907 bled from circumcision
 - Alfonso XIII blamed Ena, grew to despise her
 - More boys: Jaime, deaf-mute; Gonzalo, hemophilic
 - Spaniards blamed the British
- 1931, royal family in exile in Rome
 - Alfonso XIV was too ill to appear
 - 1933, marries Cuban commoner, abdicates
- Juan, born 1913, normal
 - His son **Juan Carlos** was King of Spain 1975 'til abdication 6/2/2014




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The Romanovs: Tsar Nicholas II, Alexandra (Alix, granddaughter of Victoria) and Family




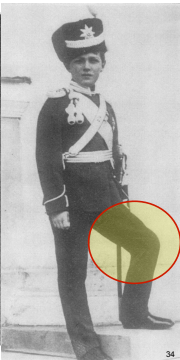
Tsarevich Alexis
b. 7/4/1904; 301 gun salute



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Prince Alexis, 1912

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Grigori Yefimovich Rasputin 1869–1916

- 1869, Born Pokrovskoye, Siberia
 - Observed two sibs drown
- 1887: 3 months in Verkhoturys monastery
- 1901: *strannik* (pilgrim), wandered Greece, Jerusalem
- 1903: Saint Petersburg, *starets* (holy man) with healing & prophetic powers
- 1905: Introduced to Alexandra by Anna Vyrubova to help with Alexis' hemophilia





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Rasputin's Power Rises 1912 Belovezhski Forest Holiday

- Alexis is near death, telegram to Rasputin in Siberia
- Calming influence, distraction?
 - "God has seen your tears and heard your prayers. Don't be sad, the little boy is not going to die. Don't let the doctors frighten him"
- Leeches?, Faith healer, hypnotism?

Rasputin became the czar's primary adviser and political gatekeeper. He engaged in monumental debauchery, and was hated by the Russian nobles, though loved as a religious figure by the Tsarina and many of the peasants.



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The Romanovs in 1912

Pierre Gilliard, Alexis' tutor, wrote: *"The illness of the Tsarevich cast its shadow over the whole of the concluding period of Tsar Nicholas II's reign. Without appearing to be, it was one of the main causes of his fall, for it made possible the phenomenon of Rasputin and resulted in the fatal isolation of the sovereigns who lived in a world apart, wholly absorbed in a tragic anxiety that had to be concealed from all eyes."*

Historians have since disputed the contribution of Alexis' hemophilia to Russian politics, but the strain on the royal household is clear enough.



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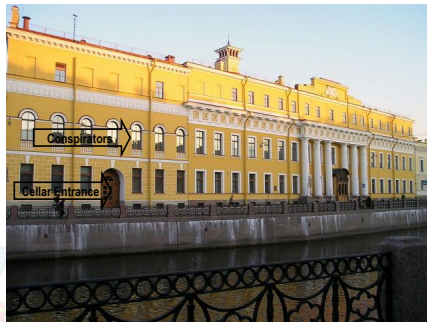
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Rasputin and Admirers, 1914


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Yusupov Palace, St. Petersburg


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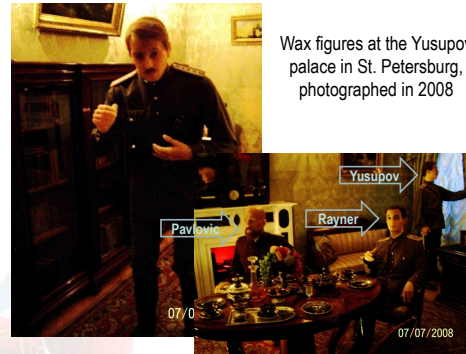
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Maj. Felix Yusupov: Dec 16, 1916

Wax figures at the Yusupov palace in St. Petersburg, photographed in 2008



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Poisoning Rasputin


Cyanide-laced cakes



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Treatment Attempts 1901–1942

- Lime
- Gelatin
- Oxygen
- Splenectomy
- Bone marrow
- Sodium citrate
- Calcium lactate
- Witte's peptone
- Hydrogen peroxide
- Induced anaphylaxis
- Antidiphtheric serum
- The 'galvanic needle'
- Animal and human sera
- Adrenaline
- Bird's muscle
- IV oxalic acid
- Vitamin therapy
- X-ray irradiation
- Serum from the mother
- Tissue fibrinogen by mouth
- Bromide extract of egg white; sedative
- Blood—both withdrawn and injected therapeutically
 - "Auto-hemotherapy"
- Female hormone therapy
 - in the belief that femininity prevents expression of the hemophilic gene

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Effective Treatments

- 1926, Surgeon General: 12 referenced attempts at whole blood transfusion beginning in WW I
- 1934, McFarlane: topical application of Russell viper venom
- 1937, Patek and Taylor first characterization of anti-hemophilic globulin
- 1938, McFarlane: fresh whole plasma
- 1950s: EJ Cohn fractionation of whole human and animal plasma
 - Animal: Biggs and Macfarlane, 1954; Bidwell, 1955
 - Kekwick and Wolf, 1957; Soulier, Gobbi, Larrieu, 1957; Blomback, Blomback, Nilsson, 1958

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1952: Stephen Christas, Canadian Hemophilia Society: *Delineation of factor IX deficiency (Christmas disease) from factor VIII deficiency (Alexis was factor IX deficient)*



1953: Nine-year-old Donald Burns smiles on father's knee. He is believed to be first successful appendectomy operation to a haemophilic in North America.

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Breakthroughs

- 1964, Judith G. Pool (1919–75, U of Chi)
 - Developed cryoprecipitate
 - First opportunity for hemophilic home care
- 1968, Kenneth M. Brinkhous
 - 1908–2000, UNC Chapel Hill
 - First to chemically characterize factor VIII in 1938
 - Developed AHF with hemophilic dog experiments
 - AHF released through Hyland in 1968







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Advances in the 1960–70s

- 1960: prevent bleeds during dental extractions and minor procedures using Kohn fraction
- 1960: surgical orthopedic correction of hemarthroses using AHF
- 1970: home therapy
- 1973: hemophilia treatment centers
- 1980: prothrombin complex concentrate
 - Proplex®, II, VII, IX, X for IX deficiency
- 1980: activated PCC, FEIBA® for inhibitors
- 1980: life expectancy was 60
 - But high hepatitis rate, 20,000 donors/pool
 - No viral inactivation

Rev. Robert K. Massie III

Robert K. Massie

Massie R, Massie K. *Journey*. Knopf, USA 1973

Susan Massie


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AIDS

- Fall 1980: *Pneumocystis carinii* pneumonia (PCP) and Kaposi sarcoma (KS) in homosexual males
 - Searched for non-infectious immunodeficiency causes such as amyl nitrite "poppers," anti-sperm antibodies or anal intercourse
- Spring 1982: CDC recorded three cases of PCP in hemophiliacs receiving AHF, all died
 - Paralleled reports of similar symptoms in Haitian hemophiliacs and in US drug abusers
 - No homosexual behavior or illegal drug use
 - Led to concept of blood-borne viral infection


Evatt BL. The tragic history of AIDS in the hemophilia population, 1982–1984. *J Thrombos Haemost* 2006; 4: 2295–301.



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July 27, 1982



- CDC reported to blood and plasma industries, gay organizers, hemophilia organizations, FDA and NIH
- Consensus to not act
 - Evidence of three patients too weak to conclude hemophiliacs were a risk group
 - Concern it would stigmatize homosexual community
 - Lose homosexual donors to blood and plasma services
 - Millions of dollars to change manufacturing policies
- Fall of 1982: six more CDC cases, including two non-hemophilic blood recipients
 - Donor services would not reveal donor identities to avoid embarrassing and subsequently losing donors
 - One definite identification

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January 4, 1983


- CDC reported the statistical prevalence of hepatitis B was identical in hemophiles and AIDS risk groups (surrogate association)
- CDC reported to the same groups including ARC, AABB, National Hemophilia Foundation, National Gay Task Force, Pharmaceutical Mfrs Association, Council of Community Blood Centers, State and Territorial Epidemiologists, and individuals.
- Again, consensus to not act, debate was irrational, acrimonious and public, harshly critical of Dr. Evatt and CDC


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January 13, 1983

- CCBC and AABB: "transfusions are life-saving procedures; some adverse reactions are acceptable to save lives. The rare disorder affecting nine cases is not enough to force a policy change."
- ARC head Dr Cumming wrote: "It has long been noted that CDC increasingly needs a major epidemic to justify its existence... In short, we can not depend on the CDC to provide scientific, objective, unbiased leadership."
- The National Hemophilia Foundation, however, alarmed, had contacted plasma manufacturers in December, 1982

 **American Red Cross**

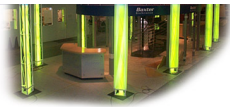

 **NATIONAL HEMOPHILIA FOUNDATION**
for all bleeding and clotting disorders

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NHF Initiative: 1983

- Dec, 1982: *Alpha Therapeutics* began to screen donors
- 20% of commercial plasma came from donor services who refused to screen donors for sexual orientation
- US Public Health Service guidelines, March 4, 1983
 - CDC bypassed FDA, sent guidelines direct to PHS
 - Donor screening and surrogate testing: hepatitis markers
- March, 1983, Baxter Hyland began heat treating plasma
- August, 1983, 26 confirmed cases of transfusion transmitted AIDS, including one F IX deficiency


 

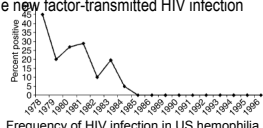
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Virus Isolation: 1983

- July, 1983: Pasteur Institute isolates virus from lymphadenopathy patients
- Feb, 1984: Pasteur Institute isolates virus from AIDS samples provided by CDC
- Sep, 1984: Alpha and Cutter demonstrate heat treatment is safe and does not raise immunogenicity
- Oct, 1984: CDC/PHS screening and heat treatment guidelines published and adopted
- By 1984, 63% of 15,500 US hemophilia patients had HIV
- Since 1/1/1985, not a single new factor-transmitted HIV infection has been recorded







Frequency of HIV infection in US hemophilia birth cohorts. (From medical records)

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Recombinant Clotting Factors

- 4/7/1976: *Genentech* incorporated
- 1981: *Genetics Institute* incorporated
- 1982: rFIX cloned by both (small molecule)
- Aug, 1984: Both cloned rFVIII gene and produced the protein
- 1985: rVWF coexpressed

Pipe SW. Recombinant clotting factors. *Thromb Haemost* 2008; 99: 840–50.

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rFVIII Barriers



- Extensive post-translational modification
 - Need for mammalian cloning system
 - Chinese hamster ovary cells
- rFVIII affinity for phospholipid
 - CHO membrane binds and degrades FVIII
 - CHO cells transfected with FVIII and VWF genes
 - Resolved by rVWF coexpression—higher affinity
- Need for serum-free cloning system
 - Eliminate animal viruses, ease purification
- High-volume fermentation process
 - Immunoaffinity, ion-exchange, Pasteurization, solvent-detergent, nanofiltration reduces 7000 L to 1 L.

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rFVIII to Market

- Regulatory uncertainty
 - Sterility, consistency questions generate 600 tests/lot
- Baxter Hyland and Genetics Institute produced Recombinate®
- Bayer Miles Lab/Cutter Biological and Genentech produced Kogenate®
- 3/27/1987: UNC Chapel Hill, first phase II infusion into a 39 year-old hemophilic
- Recombine® cleared 1992, Kogenate® 1993

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rFVIII Market Efforts and Inhibitor Formation

- B-domain deleted: ReFacto®
- No human or animal albumin in final formulation: Advate®
- Stabilizing matrix of sucrose, glycine, histidine: Kogenate FS®
- Hemostatic efficacy: 90% cessation of bleeding
- Risk of inhibitor formation in PUPs is approximately double the risk from plasma-derived FVIII
- Risk of inhibitor formation in response to rFVIII in previously treated persons (PTPs) is <1%
- 2013: Biogen Idec long half-life FVIII, Eloctate, FIX, Alprolix


Pipe SW. The promise and challenges of bioengineered recombinant clotting factors. *J Thromb Haemost* 2005; 3: 1692–1701.
Barnes C, Lillcrap D, Pazmino-Canizares J, et al: Pharmacokinetics of recombinant factor VIII (Kogenate-FS®) in children and causes of inter-patient pharmacokinetic variability. *Haemophilia* 2006;2 (Suppl 4): 40–9.

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rFIX

- 1998: Genetics Institute BeneFix®
- Barrier: γ -carboxylation of 12 glutamic acids at amino terminus
 - Requires vitamin K
 - Coexpressed γ -glutamyl carboxylase
 - Coexpression of furin
- 90% bleeding cessation using 1–2 infusions
- PUPs—3% incidence of high titer inhibitors



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Primary Prophylaxis in Children

- 2005: Joint damage outcome study
 - 25 IU/kg every other day generates 6X decrease in joint deterioration by MRI vs on-demand (OD) Rx up to 6 YO
- 2009 Italian study on prophylaxis
 - 10 Y f/u on 25 IU/kg 3X a week vs OD: 0.52 vs 1.08 total bleeds and 0.2 vs 0.52 joint bleeds/patients/month
- 2009 Danish/Russian study
 - Orthopedic issues 15.6 in OD vs 2.2 in prophylaxis

Franchini M, Coppola A, Molinari AC, et al. Forum on the role of recombinant factor VIII in children with severe haemophilia A. *Haemophilia* 2009; 1–9.
Gringeri A, Lundin V, von Mackensen S, et al. Primary and secondary prophylaxis in children with haemophilia A reduces bleeding frequency and arthropathy development compared to on demand treatment; a 10-year, randomized clinical trial. *J Thromb Haemost* 2009;7
Ingerslev J, Lethagen S, Poulsen L, et al. A case-controlled Danish-Russian comparative study of clinical outcomes in younger severe haemophilia patients treated with prophylaxis compared to those managed with on-demand treatment. *J Thromb Haemost* 2009;7

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Primary Prophylaxis in Children

- Encouragement: abundant safe rFVIII
- Barriers: venous access device clotting; cost, compliance
- Inhibitor formation in severe hemophiliacs
 - Meta-analysis of 20 trials: 1248 pts, 798 severe, 632 treated exclusively w/ pdFVIII and 616 w/ rFVIII, median age 12
 - Combined data: 14% inhibitors in pdFVIII, 25% in rFVIII
 - High titer inhibitors: 8.8% in pdFVIII, 12.3% in rFVIII

Iorio A, Marcucci M, Marchesini E, Mannucci P. Plasma derived and recombinant factor VIII concentrates include a different rate of inhibitor development in hemophilia A patients. A systematic review of the literature. *J Thromb Haemost* 2009;7

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On-demand Versus Primary Prophylaxis

- 19 hemophiliacs 30–45 YO, all <1 unit/dL
 - Mean 2 bleeds/month, no additional bleeding disorder
 - ≥ 3 bleeds in elbow, ankle or knee, prior 6 months
 - Bayer Kogenate FS: sucrose-stabilized rFVIII
 - Prospective, open-label
- Six months on-demand Rx per bleed
 - Rx 20–100 units/kg every 12–24 h to cessation
- Seven months' prophylaxis
 - Month 7 (1st month of prophylaxis) was run-in
 - Home: 20–40 units/kg 2 mL/min infusion 3x/week
- Outcomes
 - Primary: patient self-assessed joint bleeds
 - Secondary: all bleeds, joint function, quality of life, health economics and safety

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Results

| | Median of 19 subjects | 6 mo on-demand | 6 mo prophylaxis |
|--------------------------|-----------------------|----------------|------------------|
| Infusions | 29 | 88 | |
| Total consumption, units | 70,421 | 211,933 | |
| Joint bleeds | 15 | 0 | |
| All bleeds | 20.5 | 0 | |
| Spontaneous bleeds | 13.5 | 0 | |
| Trauma bleeds | 2.5 | 0 | |

- Baseline: median joint bleeds: 14.0; 24% mild, 63% moderate, 13% severe
- Fourteen used rFVIII, two used plasma-derived FVIII, three used both
- Prophylaxis median trough: 48 h: 6 units, 72 h: 4 units
- Mean total Gilbert joint function score (pain, swelling, atrophy, deformity, range of motion, instability): on-demand, 25.3%, prophylaxis 19.8%
- Safety: no Rx-related adverse events

Collins P, Faradji A, Morfini M, Enriquez MM, Schwartz L. J Thromb Haemost 2010;8:63-9. 61
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Extended Half-life Factors VIII and IX

- Recombinant B-domain deleted Fc fusion factor VIII
 - Biogen Idec Eloctate, FDA-approved 2014
 - 96 adult hemophilic males with > 12 annual bleeds, 3-4 doses/week
 - Extended by Fc receptor and IgG recycling pathway
 - rFVIII-Fc half-life 19 h versus rFVIII 12; 1.6-3.6 annual bleeds
 - Prophylactic Rx frequency up to 5 days versus 3-4 doses/week
 - Monitor using clot-based FVIII assay with kaolin-based PTT
- Recombinant Fc fusion factor IX
 - Biogen Idec Alprolix, FDA-approved 2014
 - Monitor using FIX assay with kaolin-based PTT or chromogenic assay
 - rFIX-Fc half-life 70-80 h versus 24 h, 7-10 day intervals

Shapiro AD¹, Ragni MV, Kulkarni R, et al. Recombinant factor VIII Fc fusion protein: extended-interval dosing maintains low bleeding rates and correlates with von Willebrand factor levels. J Thromb Haemost. 2014;12:1788-800.

Mancuso ME, Mannucci PM. Fc-fusion technology and recombinant FVIII and FIX in the management of the hemophilias. Drug Des Devel Ther. 2014 28;365-71. 62
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Cell-based Hemophilia Therapy

- Genetic delivery targets: liver, skeletal muscle, hematopoietic tissue, endothelial cells
- Vectors: adeno-associated virus (AAV) and liver sinusoidal endothelial cells (LSEC)
- Barriers: T-cell immune response to vector, FVIII molecule too large for vector, inefficient transduction

- Fomin ME, Togariti P, Muench MO. Progress and challenges in the development of a cell-based therapy for hemophilia A. J Thromb Haemost 2014; 12: 1954-64.
- Chuah MK, Nair N, Vandendriessche T. Recent progress in gene therapy for hemophilia. Hum Gene Ther 2012; 23:557-65.
- Lozier J. Gene therapy of the hemophilias. Semin Hematol 2004; 41:287-96.

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FIX Gene Transfer: Landmark Study

- Adenovirus-associated virus vector (AAV)
 - Codon-optimized human factor IX transgene
 - scAAV2/8-LP1-hFIXco
- Six hemophilia B (<1% FIX) patients
 - 29-64 YO, 2 with FIX null
 - Four with missense mutations (type II deficiency)
 - All receiving prophylactic FIX 3X/week
 - No inhibitors, no antibodies to AAV
- Vector administration dosages
 - 2×10^{11} vector genomes/kg body weight (vg/kg)
 - 6×10^{11} vg/kg
 - 2×10^{12} vg/kg

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FIX Transfer Study Outcomes

- 4 of 6 required no FIX Rx
- 2 required on-demand FIX Rx
 - One ran the half-marathon
- High-dose subjects had rise in ALT
 - Immune response to hepatocytes?
 - Controlled with prednisolone
- No anti-FIX antibodies
- Did not detect cellular immunity to vector

Nathwani, AC, Tuddenham GD, Rangarajan S, et al. Adenovirus-associated virus vector-mediated gene transfer in hemophilia B. NEJM 2011;365: 2357-65.

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Current Factor VIII Cellular Transfer Trial

- FVIII gene is 7 kb, AAV capacity 4.6 KB
- B-domain-reduced FVIII with minimal glycosylation
- Codon optimization: AAV8 variant has capacity to serve as a vector
- Successful in mice and macaques
- Human trials in progress

McIntosh J, Lenting PJ, Rosales C, et al. Therapeutic levels of FVIII following a single peripheral vein administration of rAAV vector encoding a novel human factor VIII variant. Blood. 2013 25;121:3335-44

Ward NJ, Buckley SM, Waddington SN, et al. Codon optimization of human factor VIII cDNAs leads to high-level expression. Blood 2011;117:798-807.

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